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<b>(54) Title:</b> AGP-1 FUSION PROTEIN COMPOSITIONS AND METHODS		
<b>(57) Abstract</b>  The present invention relates to Fc-AGP-1 fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to a genetic or chemical fusion protein comprising the Fc immunoglobulin region, derivative or analog fused to the N-terminal portion of the AGP-1 protein, derivative or analog.		

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## AGP-1 FUSION PROTEIN COMPOSITIONS AND METHODS

Field of the Invention

5           The present invention relates to AGP-1 fusion protein compositions and methods of preparation and use thereof.

Background of the Invention

10           The availability of recombinant proteins for therapeutic use has led to advances in protein modifications in order to enhance or improve the properties of such proteins as pharmaceutical agents. Such modifications can provide enhanced protein  
15 protection and decreased degradation by reducing or eliminating proteolysis. Additional advantages include, under certain circumstances, increasing the stability, circulation time, and the biological activity of the therapeutic protein. A review article  
20 describing protein modifications is Francis, *Focus on Growth Factors* 3:4-10 (May 1992) (published by Mediscript, London, UK).

          One such modification is the use of an Fc region of an immunoglobulin molecule. Antibodies  
25 comprise two functionally independent parts, a variable domain known as "Fab", which binds antigen, and a constant domain, known as "Fc" which provides the link to effector functions such as complement or phagocytic cells. The Fc portion of an immunoglobulin has a long  
30 plasma half-life, whereas the Fab is short-lived. (Capon, et al., *Nature* 337, 525-531 (1989)).

          Therapeutic protein products have been constructed using the Fc domain to provide longer half-life or to incorporate functions such as Fc receptor  
35 binding, protein A binding, complement fixation and placental transfer which all reside in the Fc proteins

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of immunoglobulins. *Id.* For example, the Fc region of an IgG1 antibody has been fused to the N-terminal end of CD30 ligand (CD30-L), a molecule which binds CD30 receptors expressed on Hodgkin's Disease tumor cells, anaplastic lymphoma cells, T-cell leukemia cells and other malignant cell types. See, U.S. Patent No. 5,480,981. IL-10, an anti-inflammatory and antirejection agent has been fused to murine Fc $\gamma$ 2a in order to increase the cytokines short circulating half-life. (Zheng et al., The Journal of Immunology, 154, 5590-5600 (1995)). Studies have also evaluated the use of tumor necrosis factor receptor linked with the Fc protein of human IgG1 to treat patients with septic shock. (Fisher et al., N. Engl. J. Med., 334: 1697-1702 (1996); Van Zee et al., The Journal of Immunology, 156: 2221-2230 (1996)). Fc has also been fused with CD4 receptor to produce a therapeutic protein for treatment of AIDS. See, Capon et al., Nature, 337:525-531 (1989). In addition, the N-terminus of interleukin-2(IL-2) has also been fused to the Fc portion of IgG1 or IgG3 to overcome the short half life of IL-2 and its systemic toxicity. See, Harvill et al., Immunotechnology, 1, 95-105 (1995).

A Type II transmembrane protein of the tumor necrosis factor (TNF) family has been identified and observed to induce apoptosis in certain tumor cell lines, but not in normal cell lines. This protein has been referred to as TRAIL (Wiley et al. Immunity 3, 673-682 (1995)), Apo-2 (Pitti et al. J. Biol. Chem. 271, 12687-12690 (1996) and AGP-1 (WO97/46686) and is referred to herein as AGP-1. AGP-1 has been studied as a soluble protein which lacks transmembrane and intracellular domains and is most advantageously used as a therapeutic in this form. However, the apoptotic



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activity of soluble AGP-1 is too low for it to be useful as a therapeutic.

Consequently, there exists a need to develop AGP-1 protein compositions for clinical application.

- 5 Such development would include AGP-1 protein compositions which achieve increased biological activity, decreased degradation, increased stability and increased circulation time. The present invention provides such compositions.

10

#### Summary of the Invention

- The present invention relates to Fc-AGP-1 fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the
- 15 present invention relates to a fusion protein comprising an Fc protein, or variant, fragment or derivative thereof, fused to the N-terminal portion of an AGP-1 protein, or variant, fragment, or derivative thereof. Unexpectedly, as described herein, a fusion
- 20 of an Fc protein to the N-terminus of a soluble AGP-1 protein demonstrates enhanced biological activity compared to an unmodified soluble AGP-1 protein. Such unexpected advantages from the Fc modification to AGP-1 protein would be advantageous in that these changes
- 25 contribute to lower doses required or less frequent dosing. Thus, as described below in more detail, the present invention has a number of aspects relating to the modification of proteins via fusion of an Fc region to an AGP-1 protein (or variant, fragments or
- 30 derivative thereof), as well as, specific modifications, preparations and methods of use thereof.

- The present invention provides for a protein having a formula selected from the group consisting of:
- 35  $R_1 - R_2$  and  $R_1 - L - R_2$ , wherein  $R_1$  is a Fc protein, or a variant or fragment thereof,  $R_2$  is an AGP-1 protein, or variant or fragment thereof, and L is a linker. The

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invention provides for genetic or chemical linkages of the R1 and R2 moieties as described herein.

In one aspect, the present invention provides a Fc-AGP-1 fusion protein wherein Fc (or a variant, fragment or derivative thereof) is genetically fused to the N-terminus of an AGP-1 protein (or a variant, fragment or derivative thereof). In another aspect of the invention, an Fc portion may also be linked to the N-terminus of an AGP-1 protein (or a variant, fragment or derivative thereof) by a peptide or chemical linker as known in the art. As noted above and described in more detail below, the Fc-AGP-1 fusion protein has unexpected enhanced biological activity when compared to a soluble AGP-1 protein. Additional aspects of the present invention, therefore, include not only Fc-AGP-1 fusion protein compositions, but also nucleic acid sequences encoding such proteins, related vectors and host cells containing such vectors, both useful for producing fusion proteins of the present invention.

In a second aspect, the present invention provides for preparing the Fc-AGP-1 fusion protein. Such methods include recombinant DNA techniques for preparation of recombinant proteins. Furthermore, such aspects include methods of protein production and purification as well.

In another aspect, the present invention provides methods for treating proliferative disorders, such as cancer or cardiovascular diseases, viral infections and viral-induced diseases, and immune disorders by administration of Fc-AGP-1 fusion proteins.

In another aspect, the present invention also provides for related pharmaceutical compositions of the Fc-AGP-1 proteins, variants, fragments and derivatives thereof, for use in the above therapies.

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Description of the Figures

Figure 1 (SEQ ID NO: 32) shows the amino acid sequence of the hinge, CH2 and CH3 regions of human IgG $\gamma$ 1.

5

Figure 2 (SEQ ID NOS: 33 and 34) shows the nucleotide and amino acid sequence of human AGP-1.

Figure 3 (SEQ ID NOS: 35 and 36) shows the nucleotide and amino acid sequence of Fc-huAGP-1 (95-281). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to human AGP-1 (95-281) are bracketed.

Figure 4 (SEQ ID NOS: 37 and 38) shows the nucleotide and amino acid sequence of Fc-huAGP-1 (114-281). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to human AGP-1 (114-281) are bracketed.

20

Figure 5 (SEQ ID NOS: 39 and 40) shows the nucleotide and amino acid sequence of Fc-muAGP-1 (99-291). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to murine AGP-1 (99-291) are bracketed.

25

Figure 6 (SEQ ID NOS: 41 and 42) shows the nucleotide and amino acid sequence of Fc-muAGP-1 (120-291). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to murine AGP-1 (120-291) are bracketed.

30

Figure 7 shows the activity of soluble AGP-1 and Fc-AGP-1 fusion protein in inducing apoptosis in cultured Jurkat cells.

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Detailed Description of the Invention

The present invention relates to Fc-AGP-1 fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to the genetic or chemical fusion of the Fc region of immunoglobulins to the N-terminal portion of the AGP-1 protein. Unexpectedly, fusion of Fc at the N-terminus of the AGP-1 protein demonstrates advantages which are not seen in soluble AGP-1 protein. Surprisingly, the N-terminally modified Fc-AGP-1 protein provides unexpected increased biological activity. Accordingly, the Fc-AGP-1 fusion protein, and variants, fragments and derivatives thereof, as well as, related methods of use and preparation, are described in more detail below.

The term "Fc" refers to a molecule or sequence comprising the sequence of a non-antigen-binding portion of antibody, whether in monomeric or multimeric form. The original immunoglobulin source of an Fc is preferably of human origin and may be from any isotype, e.g., IgG, IgA, IgM, IgE or IgD. One method of preparation of an isolated Fc molecule involves digestion of an antibody with papain to separate antigen and non-antigen binding portions of the antibody. Another method of preparation of an isolated Fc molecules is production by recombinant DNA expression followed by purification of the Fc molecules so expressed. A full-length Fc consists of the following Ig heavy chain regions: CH1, CH2 and CH3 wherein the CH1 and CH2 regions are typically connected by a flexible hinge region. In one embodiment, an Fc has the amino acid sequence of IgG1 such as that shown in Figure 1. The terms "Fc protein", "Fc sequence", "Fc molecules", "Fc region" and "Fc portion" are taken to have the same meaning as "Fc".

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The term "fragment" when used in association with Fc or AGP-1 polypeptides, or fusion polypeptides thereof, refers to a peptide or polypeptide that comprises less than the full length amino acid sequence of an Fc or AGP-1 polypeptide. Such a fragment may arise, for example, from a truncation at the amino terminus, a truncation at the carboxy terminus, and/or an internal deletion of a residue(s) from the amino acid sequence. AGP-1 or Fc fragments may result from alternative RNA splicing or from *in vivo* protease activity.

The term "variant" when used in association with Fc or AGP-1 polypeptides, or with fusion polypeptides thereof, refers to a polypeptide comprising an amino acid sequence which contain one or more amino acid sequence substitutions, deletions, and/or additions as compared to native Fc or AGP-1 polypeptide amino acid sequences. Variants may be naturally occurring or artificially constructed. Variants of the invention may be prepared from the corresponding nucleic acid molecules encoding said variants, which have a DNA sequence that varies accordingly from the DNA sequences for native Fc or AGP-1 polypeptides.

The term "derivative" when used in association with Fc or AGP-1 polypeptides, or with fusion polypeptides thereof, refers to Fc or AGP-1 polypeptide variants or fragments thereof, that have been chemically modified, as for example, by covalent attachment of one or more polymers, including, but limited to, water soluble polymers, N-linked or O-linked carbohydrates, sugars, phosphates, and/or other such molecules. The derivatives are modified in a manner that is different from native Fc or AGP-1, either in the type or location of the molecules attached to the polypeptide. Derivatives further

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includes deletion of one or more chemical groups naturally attached to an Fc or AGP-1 polypeptide.

The term "fusion" refers to joining of different peptide or protein segments by genetic or chemical methods wherein the joined ends of the peptide or protein segments may be directly adjacent to each other or may be separated by linker or spacer moieties such as amino acid residues or other linking groups.

#### 10 Compositions

The invention provides for FcAGP-1 fusion polypeptides and compositions thereof. Fusions of an Fc region to an AGP-1 polypeptide are advantageously made at the amino terminus of AGP-1, that is, the carboxy terminus of an Fc region is fused to the amino terminus of AGP-1. These fusion proteins (and nucleic acids encoding same) are designated herein as FcAGP-1. However, it is also contemplated that, in certain instances, it may be desirable to fuse the carboxy terminus of AGP-1 to the amino terminus of an Fc region. The fusion proteins (and nucleic acids encoding same) are designated herein as AGP-1Fc.

An Fc, or a variant, fragment or derivative thereof, may be from an Ig class. In one embodiment, an Fc is from the IgG class, such as IgG1, IgG2, IgG3, and IgG4. In another embodiment, an Fc is from IgG1. An Fc may also comprise amino acid residues represented by a combination of any two or more of the Ig classes, such as residues from IgG1 and IgG2, or from IgG1, IgG2 and IgG3, and so forth. In one embodiment, an Fc region of an Fc-AGP-1 fusion protein has the sequence as set forth in Figure 1 (SEQ ID NO: 32) comprising hinge CH2 and CH3 regions of human IgG1. (see Ellison et al., Nucleic Acids Res. 10, 4071-4079 (1982)).

In addition to naturally occurring variations in Fc regions, Fc variants, fragments and derivatives

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may contain non-naturally occurring changes in Fc which are constructed by, for example, introducing substitutions, additions, insertions or deletions of residues or sequences in a native or naturally occurring Fc, or by modifying the Fc portion by chemical modification and the like. In general, Fc variants, fragments and derivatives are prepared such that the increased circulating half-life of Fc fusions to AGP-1 is largely retained.

Also provided by the invention are Fc variants with conservative amino acid substitutions. The term "conservative amino acid substitution" refers to a substitution of a native amino acid residue with a nonnative residue such that there is little or no effect on the polarity or charge of the amino acid residue at that position. For example, a conservative substitution results from the replacement of a non-polar residue in a polypeptide with any other non-polar residue. General rules for conservative amino acid substitutions are set forth in Table I.

Table I  
Conservative Amino Acid Substitutions

Original Residues	Exemplary Substitutions	Preferred Substitutions
Ala	Val, Leu, Ile	Val
Arg	Lys, Gln, Asn	Lys
Asn	Gln, His, Lys, Arg	Gln
Asp	Glu	Glu
Cys	Ser	Ser
Gln	Asn	Asn
Glu	Asp	Asp
Gly	Pro, Ala	Ala
His	Asn, Gln, Lys, Arg	Arg
Ile	Leu, Val, Met, Ala, Phe, Norleucine	Leu

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Leu	Norleucine, Ile, Val, Met, Ala, Phe	Ile
Lys	Arg, Gln, Asn	Arg
Met	Leu, Phe, Ile	Leu
Phe	Leu, Val, Ile, Ala, Tyr	Leu
Pro	Ala	Ala
Ser	Thr	Thr
Thr	Ser	Ser
Trp	Tyr, Phe	Tyr
Tyr	Trp, Phe, Thr, Ser	Phe
Val	Ile, Met, Leu, Phe, Ala, Norleucine	Leu

Conservative amino acid substitutions also encompass non-naturally occurring amino acid residues which are typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems. These include peptidomimetics, and other reversed or inverted forms of amino acid moieties. Conservative modifications to the amino acid sequence (and the corresponding modifications to the encoding nucleotides) are expected to produce Fc molecules (and FcAGP-1 fusion proteins) having functional and chemical characteristics similar to those of unmodified Fc and FcAGP-1 proteins.

In addition to the substitutions set forth in Table I, any native residue in an Fc region (or in an FcAGP-1 fusion protein) may also be substituted with alanine, as has been previously described for "alanine scanning mutagenesis" (Cunningham et al. Science 244, 1081-1085 (1989)).

Substantial modifications in the functional and/or chemical characteristics of an Fc molecule (and an FcAGP-1 fusion protein) may be accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the molecular backbone in the area of the substitution, for



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example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues may be divided into groups based on

5 common side chain properties:

- 1) hydrophobic: norleucine, Met, Ala, Val, Leu, Ile;
- 2) neutral hydrophilic: Cys, Ser, Thr;
- 3) acidic: Asp, Glu;
- 10 4) basic: Asn, Gln, His, Lys, Arg;
- 5) residues that influence chain orientation: Gly, Pro; and
- 6) aromatic: Trp, Tyr, Phe.

Non-conservative substitutions may involve  
15 the exchange of a member of one of these classes for a member from another class. Such substituted residues may be introduced into regions of an Fc or AGP-1 molecule that are homologous with non-human Fc or AGP-1, or into the non-homologous regions of the molecule.

20 Cysteine residues in Fc molecules can be deleted or replaced with other amino acids to prevent formation of disulfide crosslinks. In particular, a cysteine residue at position 5 of Figure 1 (SEQ. ID. NO. 32) may be substituted with one or more amino  
25 acids, such as alanine or serine. Alternatively, the cysteine residue at position 5 could be deleted.

An Fc fragment may be prepared by deletion of one or more amino acids at any of positions 1, 2, 3, 4 and 5 as shown in Figure 1 (SEQ ID NO. 32). In one  
30 embodiment, the amino acid residues at positions 1-5 inclusive are deleted. Substitutions at these positions can also be made and are within the scope of this invention.

Fc variants may also be made which show  
35 reduced binding to Fc receptors which trigger effector functions such as antibody dependent cellular

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cytotoxicity (ADCC) and activation of complement. Such variants may include leucine at position 20 deleted or substituted with a glutamine residue, glutamate at position 103 deleted or substituted with an alanine residue, and lysines at positions 105 and 107 deleted or substituted with alanine residues (following the numbering as set forth in Figure 1). One or more of such substitutions are contemplated.

In one embodiment, Fc variants will exhibit stronger binding to the FcRn receptor ("salvage receptor") and a longer circulating half-life compared to native Fc such as that shown in Figure 1. Example of such variants include amino acid substitutions at one or more of residues 33, 35-42, 59, 72, 75, 77, 95-98, 101, 172-174, 215 and 220-223, wherein the substitution(s) confer tighter binding of an Fc variant to the FcRn receptor.

Other Fc variants include one or more tyrosine residues replaced with, for example, phenylalanine residues. In addition, other variant amino acid insertions, deletions and/or substitutions are also contemplated and are within the scope of the present invention. Examples include Fc variants disclosed in WO96/32478 and WO97/34630 hereby incorporated by reference. Furthermore, alterations may be in the form of altered amino acids, such as peptidomimetics or D-amino acids.

The Fc protein may be also linked to the AGP-1 proteins of the Fc-AGP-1 protein by "linker" moieties whether chemical or amino acids of varying lengths. Such chemical linkers are well known in the art. Amino acid linker sequences can include but are not limited to:

- (a) ala-ala-ala;
- (b) ala-ala-ala-ala;
- (c) ala-ala-ala-ala-ala;

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- (d) gly-gly;  
(e) gly-gly-gly;  
(f) gly-gly-gly-gly-gly;  
(g) gly-gly-gly-gly-gly-gly-gly;  
5 (h) gly-pro-gly;  
(i) gly-gly-pro-gly-gly; and  
(j) any combination of subparts (a) through (i).

10 AGP-1 variants, fragments and derivatives are also provided by the invention and are generally as described hereinabove for Fc molecules, with the exception of the specific locations of the modified amino acid residues. In a preferred embodiment, AGP-1  
15 is a soluble form of AGP-1 which is not membrane-bound and lacks a functional transmembrane domain. As an example, soluble AGP-1 may comprise an extracellular domain and lack sequences for cytoplasmic and transmembrane domains. The full-length human AGP-1  
20 extracellular domain encompasses about residues 39-281 inclusive using the numbering system as set forth in Figure 2. Soluble human AGP-1 may also encompass fragments of a full-length extracellular domain which function to bind receptor or to induce apoptosis in an  
25 assay such as that described in Example 4 below. Soluble AGP-1 fragments comprise the amino acid sequence X-281 wherein X is any residue from 95 to 114 inclusive using the numbering system of as set forth in Figure 2. Other soluble AGP-1 fragments encompass  
30 residues 115-281, 116-281, 117-281, 118-281 and 119-281 inclusive. Variants and derivatives of the AGP-1 fragments described herein are also encompassed by the invention.

35 Nucleic acid molecules

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Nucleic acid molecules encoding Fc-AGP-1 proteins, or variants, fragments or derivatives thereof, are provided for by the invention. Nucleic acid molecules of the invention may be produced using  
5 site directed mutagenesis, PCR amplification, or other appropriate methods, where the primer(s) have the desired mutations. See Sambrook et al. (Molecular Cloning: A Laboratory Manual, Cold Springs Harbor Laboratory Press, Cold Springs Harbor, N.Y. (1989)),  
10 and Ausubel et al. (Current Protocols in Molecular Biology, Wiley and Sons, N.Y. (1994)), for descriptions of mutagenesis techniques. Chemical synthesis using methods described by Engels et al. (Angew. Chem. Intl. Ed. 28, 716-734 (1989)), may also be used to prepare  
15 such variants. Other methods known to the skilled artisan may be used as well.

In certain embodiments, nucleic acid variants contain codons which have been altered for optimal expression of an Fc-AGP-1 polypeptide in a given host  
20 cell. Particular codon alterations will depend upon the Fc-AGP-1 polypeptide(s) and host cell(s) selected for expression. Such "codon optimization" can be carried out by a variety of methods, for example, by selecting codons which are preferred for use in highly  
25 expressed genes in a given host cell. Computer algorithms which incorporate codon frequency tables such as "Ecohigh. Cod" for codon preference of highly expressed bacterial genes may be used and are provided by the University of Wisconsin Package Version 9.0,  
30 Genetics Computer Group, Madison, WI. Other useful codon frequency tables include "Celegans\_high.cod", "Celegans\_low.cod", "Drosophila\_high.cod", "Human\_high.cod", "Maize\_high.cod", and "Yeast\_high.cod".

35 In other embodiments, nucleic acid molecules encode Fc-AGP-1 variants with conservative amino acid

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substitutions as defined hereinabove. Also provided for are Fc or AGP-1 variants comprising an addition and/or a deletion of one or more N-linked or O-linked glycosylation sites, or Fc or AGP-1 polypeptide fragments as described above. In addition, nucleic acid molecules may encode any combination of Fc and/or AGP-1 variants, fragments, and fusion polypeptides described herein.

#### 10 Vectors and Host cells

A nucleic acid molecule encoding an Fc-AGP-1 fusion protein is inserted into an appropriate expression vector using standard ligation techniques. The vector is typically selected to be functional in the particular host cell employed (i.e., the vector is compatible with the host cell machinery such that amplification of the gene and/or expression of the gene can occur). A nucleic acid molecule encoding an Fc-AGP-1 protein may be amplified/expressed in prokaryotic, yeast, insect (baculovirus systems) and/or eukaryotic host cells. Selection of the host cell will depend in part on whether an Fc-AGP-1 protein is to be post-translationally modified (e.g, glycosylated and/or phosphorylated). If so, yeast, insect, or mammalian host cells are preferable.

Typically, expression vectors used in any of the host cells will contain sequences for plasmid maintenance and for cloning and expression of exogenous nucleotide sequences. such sequences, collectively referred to as "flanking sequences" in certain embodiments will typically include one or more of the following nucleotides: a promoter, one or more enhancer sequences, an origin of replication, a transcriptional termination sequence, a complete intron sequence containing a donor and acceptor splice site, a leader sequence for secretion, a ribosome binding site,

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a polyadenylation sequence, a polylinker region for inserting the nucleic acid encoding the polypeptide to be expressed, and a selectable marker element.

Flanking sequences may be homologous (i.e.,  
5 from the same species and/or strain as the host cell), heterologous (i.e., from a species other than the host cell species or strain), hybrid (i.e., a combination of flanking sequences from more than one source), or synthetic, or native sequences which normally function  
10 to regulate AGP-1 and/or Fc protein expression. As such, the source of flanking sequences may be any prokaryotic or eukaryotic organism, any vertebrate or invertebrate organism, or any plant, provided that the flanking sequences is functional in, and can be  
15 activated by, the host cell machinery.

A leader, or signal, sequence may be used to direct an Fc-AGP-1 polypeptide out of the host cell. Typically, the signal sequence is positioned in the coding region of the Fc-AGP-1 nucleic acid molecule, or  
20 directly at the 5' end of the Fc-AGP-1 polypeptide coding region. Many signal sequences have been identified, and any of them that are functional in the selected host cell may be used in conjunction with nucleic acid sequences encoding Fc-AGP-1 proteins.  
25 Therefore, a signal sequence may be homologous (naturally occurring) or heterologous to the AGP-1 or Fc gene or cDNA. Additionally, a signal sequence may be chemically synthesized using methods set forth above. In most cases, secretion of an Fc-AGP-1 polypeptide  
30 from the host cell via the presence of a signal peptide will result in the removal of the signal peptide from the fusion polypeptide.

The signal sequence may be a component of the vector, or it may be a part of Fc-AGP-1 DNA that is  
35 inserted into the vector. Native AGP-1 DNA encodes a signal sequence at the amino terminus of the protein

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that is cleaved during post-translational processing of the molecule to form the mature protein (see Figure 2). Included within the scope of this invention are AGP-1 nucleotides with the native signal sequence as well as  
5 AGP-1 nucleotides wherein the native signal sequence is deleted and replaced with a heterologous signal sequence. A heterologous signal sequence selected should be one that is recognized and processed, i.e., cleaved by a signal peptidase, by the host cell. In  
10 one embodiment, a heterologous signal sequence is the OPG signal sequence as described in WO97/23614. For prokaryotic host cells that do not recognize and process the native AGP-1 signal sequence, the signal sequence is substituted by a prokaryotic signal  
15 sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, or heat-stable enterotoxin II leaders. For yeast secretion, the native AGP-1 signal sequence may be substituted by the yeast invertase, alpha factor, or acid phosphatase  
20 leaders. In mammalian cell expression the native signal sequence is satisfactory, although other mammalian signal sequences may be suitable.

Preferred vectors for practicing this invention are those which are compatible with  
25 bacterial, insect, and mammalian host cells. Such vectors include, *inter alia*, pCRII, pCR3, and pCDNA3.1 (Invitrogen Company, San Diego, CA), pBSII (Stratagene Company, La Jolla, CA), pET15b (Novagen, Madison, WI), pGEX (Pharmacia Biotech, Piscataway, NJ), pEGFP-N2  
30 (Clontech, Palo Alto, CA), pETL (BlueBacII; Invitrogen), pDSR-alpha (PCT Publication No. WO90/14363) and pFastBacDual (Gibco/BRL, Grand Island, NY).

Additional possible vectors include, but are  
35 not limited to, cosmids, plasmids or modified viruses, but the vector system must be compatible with the

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selected host cell. Such vectors include, but are not limited to plasmids such as Bluescript<sup>®</sup> plasmid derivatives (a high copy number ColE1-based phagemid, Stratagene Cloning Systems Inc., La Jolla CA), PCR cloning plasmids designed for cloning Taq-amplified PCR products (e.g., TOPO<sup>™</sup> TA Cloning<sup>®</sup> Kit, PCR2.1<sup>®</sup> plasmid derivatives, Invitrogen, Carlsbad, CA), and mammalian, yeast or virus vectors such as a baculovirus expression system (pBacPAK plasmid derivatives, Clontech, Palo Alto, CA). The recombinant molecules can be introduced into host cells via transformation, transfection, infection, electroporation, or other known techniques. After the vector has been constructed and a nucleic acid molecule encoding an AGP-1 polypeptide has been inserted into the proper site of the vector, the completed vector may be inserted into a suitable host cell for amplification and/or polypeptide expression.

Host cells may be prokaryotic host cells (such as *E. coli*) or eukaryotic host cells (such as a yeast cell, an insect cell, or a vertebrate cell). The host cell, when cultured under appropriate conditions, synthesizes an AGP-1 polypeptide which can subsequently be collected from the culture medium (if the host cell secretes it into the medium) or directly from the host cell producing it (if it is not secreted). Selection of an appropriate host cell will depend upon various factors, such as desired expression levels, polypeptide modifications that are desirable or necessary for activity, such as glycosylation or phosphorylation, and ease of folding into a biologically active molecule.

Suitable host cells or cell lines may be mammalian cells, such as Chinese hamster ovary cells (CHO) (ATCC #CCL61 and Urlaub et al., Proc. Natl. Acad. Sci. USA 77, 4216-4220 (1980)), human embryonic kidney (HEK) 293 or 293T cells (ATCC #CRL1573), or 3T3 cells



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(ATCC #CRL1658). The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening and product production and purification are known in the art. Other suitable  
5 mammalian cell lines, are the monkey COS-1 and COS-7 cell lines (ATCC #CRL1651), and the CV-1 cell line (ATCC #CCL70). Further exemplary mammalian host cells include primate cell lines and rodent cell lines, including transformed cell lines. Normal diploid  
10 cells, cell strains derived from *in vitro* culture of primary tissue, as well as primary explants, are also suitable. Candidate cells may be genotypically deficient in the selection gene, or may contain a dominantly acting selection gene. Other suitable  
15 mammalian cell lines include but are not limited to, mouse neuroblastoma N2A cells, HeLa, mouse L-929 cells, 3T3 lines derived from Swiss, Balb-c or NIH mice, BHK or HaK hamster cell lines. Each of these cell lines is known by and available to those skilled in the art.

20 Similarly useful as host cells suitable for the present invention are bacterial cells. For example, the various strains of *E. coli* (e.g., HB101, DH5a, DH10, and MC1061) are well-known as host cells in the field of biotechnology. Various strains of *B. subtilis*, *Pseudomonas spp.*, other *Bacillus spp.*,  
25 *Streptomyces spp.*, and the like may also be employed in this method.

Many strains of yeast cells known to those skilled in the art are also available as host cells for  
30 expression of the polypeptides of the present invention. Preferred yeast cells include, for example, *Saccharomyces cerevisiae*.

Additionally, where desired, insect cell systems may be utilized in the methods of the present  
35 invention. Such systems are described for example in Kitts et al. (*Biotechniques*, 14, 810-817 (1993)),

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Lucklow (*Curr. Opin. Biotechnol.*, 4, 564-572 (1993)) and Lucklow et al. (*J. Virol.*, 67, 4566-4579 (1993)). Preferred insect cells are Sf-9 and Hi5 (Invitrogen, Carlsbad, CA).

5                   Transformation or transfection of an expression vector for an AGP-1 polypeptide into a selected host cell may be accomplished by well known methods including methods such as calcium chloride, electroporation, microinjection, lipofection or the  
10 DEAE-dextran method. The method selected will in part be a function of the type of host cell to be used. These methods and other suitable methods are well known to the skilled artisan, and are set forth, for example, in Sambrook et al., *supra*.

15

#### Polypeptide Production

Host cells comprising an AGP-1 expression vector (*i.e.*, transformed or transfected) may be cultured using standard media well known to the skilled  
20 artisan. The media will usually contain all nutrients necessary for the growth and survival of the cells. Suitable media for culturing *E. coli* cells are for example, Luria Broth (LB) and/or Terrific Broth (TB). Suitable media for culturing eukaryotic cells are RPMI  
25 1640, MEM, DMEM, all of which may be supplemented with serum and/or growth factors as required by the particular cell line being cultured. A suitable medium for insect cultures is Grace's medium supplemented with yeastolate, lactalbumin hydrolysate, and/or fetal calf  
30 serum as necessary (Gibco Life Technologies, Gaithersburg, MD).

Typically, an antibiotic or other compound useful for selective growth of transfected or transformed cells is added as a supplement to the  
35 media. The compound to be used will be dictated by the selectable marker element present on the plasmid with

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which the host cell was transformed. For example, where the selectable marker element is kanamycin resistance, the compound added to the culture medium will be kanamycin; where the selectable marker element is ampicillin resistance, the compound added to the culture medium will be ampicillin.

The amount of an AGP-1 polypeptide produced by a host cell can be evaluated using standard methods known in the art. Such methods include, without limitation, Western blot analysis, SDS-polyacrylamide gel electrophoresis, non-denaturing gel electrophoresis, HPLC separation, immunoprecipitation, and/or activity assays such as DNA binding gel shift assays.

Where an AGP-1 polypeptide is prepared without a tag attached, and no antibodies are available, other well known procedures for purification can be used. Such procedures include, without limitation, ion exchange chromatography, molecular sieve chromatography, HPLC, native gel electrophoresis in combination with gel elution, and preparative isoelectric focusing ("Isoprime" machine/technique, Hoefer Scientific). In some cases, two or more of these techniques may be combined to achieve increased purity.

If an AGP-1 polypeptide is produced intracellularly, the intracellular material (including inclusion bodies for gram-negative bacteria) can be extracted from the host cell using any standard technique known to the skilled artisan. For example, the host cells can be lysed to release the contents of the periplasm/cytoplasm by French press, homogenization, and/or sonication followed by centrifugation.

If an AGP-1 polypeptide has formed inclusion bodies in the cytosol, the inclusion bodies can often

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bind to the inner and/or outer cellular membranes and thus will be found primarily in the pellet material after centrifugation. The pellet material can then be treated at pH extremes or with chaotropic agent such as a detergent, guanidine, guanidine derivatives, urea, or urea derivatives in the presence of a reducing agent such as dithiothreitol at alkaline pH or tris carboxyethyl phosphine at acid pH to release, break apart, and solubilize the inclusion bodies. An AGP-1 polypeptide in its now soluble form can then be analyzed using gel electrophoresis, immunoprecipitation or the like. If it is desired to isolate an AGP-1 polypeptide, isolation may be accomplished using standard methods such as those set forth below and in Marston et al. (Meth. Enz., 182, 264-275 (1990)).

In some cases, an AGP-1 polypeptide may not be biologically active upon isolation. Various methods for "refolding" or converting the polypeptide to its tertiary structure and generating disulfide linkages, can be used to restore biological activity. Such methods include exposing the solubilized polypeptide to a pH usually above 7 and in the presence of a particular concentration of a chaotrope. The selection of chaotrope is very similar to the choices used for inclusion body solubilization, but usually the chaotrope is used at a lower concentration and is not necessarily the same as chaotropes used for the solubilization. In most cases the refolding/oxidation solution will also contain a reducing agent or the reducing agent plus its oxidized form in a specific ratio to generate a particular redox potential allowing for disulfide shuffling to occur in the formation of the protein's cysteine bridge(s). Some of the commonly used redox couples include cysteine/cystamine, glutathione (GSH)/dithiobis GSH, cupric chloride, dithiothreitol (DTT)/dithiane DTT, and 2-

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mercaptoethanol(bME)/dithio-b(ME). In many instances,  
a cosolvent may be used or may be needed to increase  
the efficiency of the refolding and the more common  
reagents used for this purpose include glycerol,  
5 polyethylene glycol of various molecular weights,  
arginine and the like.

#### Derivatives

The present Fc-AGP-1 fusion proteins, and  
10 variants and fragments thereof, are derivatized by the  
attachment of one or more chemical moieties to an  
Fc-AGP-1 fusion protein moiety. These chemically  
modified derivatives may be further formulated for  
intraarterial, intraperitoneal, intramuscular  
15 subcutaneous, intravenous, oral, nasal, pulmonary,  
topical or other routes of administration as discussed  
below. Chemical modification of biologically active  
proteins has been found to provide additional  
advantages under certain circumstances, such as  
20 increasing the stability and circulation time of the  
therapeutic protein and decreasing immunogenicity.  
See, U.S. Patent No. 4,179,337. For a review, see  
Abuchowski et al., in Enzymes as Drugs. (J. S.  
Holcerberg and J. Roberts, eds. pp. 367-383 (1981));  
25 Francis et al., supra.

The chemical moieties suitable for such  
derivatization may be selected from among various water  
soluble polymers. The polymer selected should be water  
soluble so that the protein to which it is attached  
30 does not precipitate in an aqueous environment, such as  
a physiological environment. Preferably, for  
therapeutic use of the end-product preparation, the  
polymer will be pharmaceutically acceptable. One  
skilled in the art will be able to select the desired  
35 polymer based on such considerations as whether the  
polymer/protein conjugate will be used therapeutically,

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and if so, the desired dosage, circulation time, resistance to proteolysis, and other considerations. For the present proteins, the effectiveness of the derivatization may be ascertained by administering the derivative, in the desired form (i.e., by osmotic pump, or, more preferably, by injection or infusion, or, further formulated for oral, pulmonary or nasal delivery, for example), and observing biological effects as described herein.

10           The water soluble polymer may be selected from the group consisting of, for example, polyethylene glycol, copolymers of ethylene glycol/propylene glycol, carboxymethylcellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone, poly-1, 3-dioxolane, 15 poly-1,3,6-trioxane, ethylene/maleic anhydride copolymer, polyaminoacids (either homopolymers or random copolymers), and dextran or poly(n-vinyl pyrrolidone)polyethylene glycol, propylene glycol homopolymers, polypropylene oxide/ethylene oxide 20 co-polymers, polyoxyethylated polyols and polyvinyl alcohol. Polyethylene glycol propionaldehyde may have advantages in manufacturing due to its stability in water. Also, succinate and styrene may also be used.

          Fc-AGP-1 fusion proteins, variants and 25 fragments may also be derivatized by attaching polyaminoacids or branch point amino acids to the Fc or AGP-1 protein (or variant or fragment) moiety. For example, the polyaminoacid may be an additional carrier protein which serves to further increase the 30 circulation half life of an Fc-AGP-1 fusion protein in addition to the advantages achieved via the Fc-AGP-1 fusion protein above. For the present therapeutic or cosmetic purpose of the present invention, such polyaminoacids should be those which have or do not 35 create neutralizing antigenic response, or other adverse responses. Such polyaminoacids may be selected

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from the group consisting of serum album (such as human serum albumin), or other polyaminoacids, e.g. lysines. As indicated below, the location of attachment of the polyaminoacid may be at the N-terminus of the Fc-AGP-1 protein moiety, or C-terminus, or other places in between, and also may be connected by a chemical "linker" moiety to the Fc-AGP-1 protein.

The polymer may be of any molecular weight, and may be branched or unbranched. For polyethylene glycol, the preferred molecular weight is between about 2 kDa and about 100 kDa (the term "about" indicating that in preparations of polyethylene glycol, some molecules will weigh more, some less, than the stated molecular weight) for ease in handling and manufacturing. Other sizes may be used, depending on the desired therapeutic profile (e.g., the duration of sustained release desired, the effects, if any on biological activity, the ease in handling, the degree or lack of antigenicity and other known effects of the polyethylene glycol to a therapeutic protein or analog).

The number of polymer molecules so attached may vary, and one skilled in the art will be able to ascertain the effect on function. One may mono-derivatize, or may provide for a di-, tri-, tetra- or some combination of derivatization, with the same or different chemical moieties (e.g., polymers, such as different weights of polyethylene glycols). The proportion of polymer molecules to protein (or peptide) molecules will vary, as will their concentrations in the reaction mixture. In general, the optimum ratio (in terms of efficiency of reaction in that there is no excess unreacted protein or polymer) will be determined by factors such as the desired degree of derivatization (e.g., mono, di-, tri-, etc.), the molecular weight of

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the polymer selected, whether the polymer is branched or unbranched, and the reaction conditions.

The chemical moieties should be attached to the protein with consideration of effects on functional or antigenic domains of the protein. There are a number of attachment methods available to those skilled in the art. *E.g.*, EP 0401384 herein incorporated by reference (coupling PEG to G-CSF), see also Malik et al., Exp. Hematol. 20, 1028-1035 (1992) (reporting pegylation of GM-CSF using tresyl chloride). For example, polyethylene glycol may be covalently bound through amino acid residues via a reactive group, such as, a free amino or carboxyl group. Reactive groups are those to which an activated polyethylene glycol molecule may be bound. The amino acid residues having a free amino group may include lysine residues and the N-terminal amino acid residue. Those having a free carboxyl group may include aspartic acid residues, glutamic acid residues, and the C-terminal amino acid residue. Sulfhydryl groups may also be used as a reactive group for attaching the polyethylene glycol molecule(s). Preferred for therapeutic purposes is attachment at an amino group, such as attachment at the N-terminus or lysine group. Attachment at residues important for receptor binding should be avoided if receptor binding is desired.

One may specifically desire N-terminally chemically modified Fc-AGP-1 fusion protein. Using polyethylene glycol as an illustration of the present compositions, one may select from a variety of polyethylene glycol molecules (by molecular weight, branching, etc.), the proportion of polyethylene glycol molecules to protein (or peptide) molecules in the reaction mix, the type of pegylation reaction to be performed, and the method of obtaining the selected N-terminally pegylated protein. The method of



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obtaining the N-terminally pegylated preparation (i.e., separating this moiety from other monopegylated moieties if necessary) may be by purification of the N-terminally pegylated material from a population of pegylated protein molecules. Selective N-terminal chemical modification may be accomplished by reductive alkylation which exploits differential reactivity of different types of primary amino groups (lysine versus the N-terminal) available for derivatization in a particular protein. Under the appropriate reaction conditions, substantially selective derivatization of the protein at the N-terminus with a carbonyl group containing polymer is achieved. For example, one may selectively N-terminally pegylate the protein by performing the reaction at a pH which allows one to take advantage of the  $pK_a$  differences between the  $\epsilon$ -amino group of the lysine residues and that of the  $\alpha$ -amino group of the N-terminal residue of the protein. By such selective derivatization, attachment of a water soluble polymer to a protein is controlled: the conjugation with the polymer takes place predominantly at the N-terminus of the protein and no significant modification of other reactive groups, such as the lysine side chain amino groups, occurs. Using reductive alkylation, the water soluble polymer may be of the type described above, and should have a single reactive aldehyde for coupling to the protein. Polyethylene glycol propionaldehyde, containing a single reactive aldehyde, may be used.

30           An N-terminally monopegylated derivative is preferred for ease in production of a therapeutic. N-terminal pegylation ensures a homogenous product as characterization of the product is simplified relative to di-, tri- or other multi-pegylated products. The use of the above reductive alkylation process for

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preparation of an N-terminal product is preferred for ease in commercial manufacturing.

#### Uses of the Polypeptide

5           AGP-1 fusion proteins may be used for the treatment of proliferative disorders wherein cells are undergoing excessive proliferation. For example, an AGP-1 fusion protein may be used as an anti-tumor to treat patients suffering from a variety of cancers, 10 such as breast cancer, prostate cancer, lung cancer, and colon cancer. Viral infections and viral-induced diseases, such as hepatitis and AIDS, may also be treated with the proteins of the invention. Cardiovascular diseases such as arteriosclerosis which 15 characterized by excessive proliferation of vascular smooth muscle cells may also be treated. AGP-1 fusion proteins may also be used to suppress T-lymphocyte mediated immune responses that occur in autoimmune disorders and in rejection of transplanted tissues. It 20 is desirable that that present proteins are expected to have prolonged in vivo half-lives and circulation times that will allow for lower dosages, less frequent administration, and enhanced efficacy compared to an unfused AGP-1 protein.

25

#### Pharmaceutical Compositions

The present invention also provides for pharmaceutical compositions of the Fc-AGP-1 fusion proteins, variants, fragments and derivatives. Such 30 pharmaceutical compositions may be for administration for injection, or for oral, pulmonary, nasal, transdermal or other forms of administration. In general, comprehended by the invention are pharmaceutical compositions comprising effective 35 amounts of protein or derivative products of the invention together with pharmaceutically acceptable

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diluents, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. An effective or a therapeutically effective amount of an FcAGP-1 fusion protein is an amount sufficient to induce apoptosis in a target cell, wherein apoptosis is evaluated by assays known in the art.

Such compositions include diluents of various buffer content (e.g., Tris-HCl, acetate, phosphate), pH and ionic strength; additives such as detergents and solubilizing agents (e.g., Tween 80, Polysorbate 80), anti-oxidants (e.g., ascorbic acid, sodium metabisulfite), preservatives (e.g., Thimersol, benzyl alcohol) and bulking substances (e.g., lactose, mannitol); incorporation of the material into particulate preparations of polymeric compounds such as polylactic acid, polyglycolic acid, etc. or into liposomes. Hylauronic acid may also be used, and this may have the effect of promoting sustained duration in the circulation. Such compositions may influence the physical state, stability, rate of in vivo release, and rate of in vivo clearance of the present proteins and derivatives. See, e.g., Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712 which are herein incorporated by reference. The compositions may be prepared in liquid form, or may be in dried powder, such as lyophilized form. Implantable sustained release formulations are also contemplated, as are transdermal formulations.

Contemplated for use herein are oral solid dosage forms, which are described generally in Remington's Pharmaceutical Sciences, 18th Ed. 1990 (Mack Publishing Co. Easton PA 18042) at Chapter 89, which is herein incorporated by reference. Solid dosage forms include tablets, capsules, pills, troches

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or lozenges, cachets or pellets. Also, liposomal or proteinoid encapsulation may be used to formulate the present compositions (as, for example, proteinoid microspheres reported in U.S. Patent No. 4,925,673).

5 Liposomal encapsulation may be used and the liposomes may be derivatized with various polymers (e.g., U.S. Patent No. 5,013,556). A description of possible solid dosage forms for the therapeutic is given by Marshall, K. In: *Modern Pharmaceuticals* Edited by G. S. Banker and

10 C. T. Rhodes Chapter 10, 1979, herein incorporated by reference. In general, the formulation will include the Fc-AGP-1 fusion protein (or analog or derivative), and inert ingredients which allow for protection against the stomach environment, and release of the

15 biologically active material in the intestine.

Also specifically contemplated are oral dosage forms of the above derivatized proteins. FcAGP-1 fusion protein may be chemically modified so that oral delivery of the derivative is efficacious. Generally,

20 the chemical modification contemplated is the attachment of at least one moiety to the protein (or peptide) molecule itself, where said moiety permits (a) inhibition of proteolysis; and (b) uptake into the blood stream from the stomach or intestine. Also

25 desired is the increase in overall stability of the protein and increase in circulation time in the body. Examples of such moieties include: Polyethylene glycol, copolymers of ethylene glycol and propylene glycol, carboxymethyl cellulose, dextran, polyvinyl

30 alcohol, polyvinyl pyrrolidone and polyproline. Abuchowski and Davis, Soluble Polymer-Enzyme Adducts. In: "Enzymes as Drugs", Hoenberg and RAGP-1erts, eds., Wiley-Interscience, New York, NY, (1981), pp 367-383; Newmark, et al., J. Appl. Biochem. 4: 185-189 (1982).

35 Other polymers that could be used are poly-1,3-dioxolane and poly-1,3,6-tioxocane. Preferred for

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pharmaceutical usage, as indicated above, are polyethylene glycol moieties.

To ensure resistance to degradation in the stomach following oral administration, a coating  
5 impermeable to at least pH 5.0 is essential. Examples of the more common inert ingredients that are used as enteric coatings for oral formulations are cellulose acetate trimellitate (CAT), hydroxypropylmethylcellulose phthalate (HPMCP), HPMCP  
10 50, HPMCP 55, polyvinyl acetate phthalate (PVAP), Eudragit L30D, Aquateric, cellulose acetate phthalate (CAP), Eudragit L, Eudragit S, and Shellac. These coatings may be used as mixed films.

The therapeutic can be included in the  
15 formulation as fine multiparticulates in the form of granules or pellets of particle size about 1 mm. The formulation of the material for capsule administration could also be as a powder, lightly compressed plugs or even as tablets. The therapeutic could be prepared by  
20 compression.

One may dilute or increase the volume of an FcAGP-1 composition with an inert material. These diluents could include carbohydrates, especially mannitol,  $\alpha$ -lactose, anhydrous lactose, cellulose,  
25 sucrose, modified dextrans and starch. Certain inorganic salts may be also be used as fillers including calcium triphosphate, magnesium carbonate and sodium chloride. Some commercially available diluents are Fast-Flo, Emdex, STA-Rx 1500, Emcompress and  
30 Avicell.

Disintegrants may be included in the formulation of the therapeutic into a solid dosage form. Materials used as disintegrates include but are not limited to starch including the commercial  
35 disintegrant based on starch, Explotab. Sodium starch glycolate, Amberlite, sodium carboxymethylcellulose,

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ultramylopectin, sodium alginate, gelatin, orange peel, acid carboxymethyl cellulose, natural sponge and bentonite may all be used. Another form of the disintegrants are the insoluble cationic exchange  
5 resins. Powdered gums may be used as disintegrants and as binders and these can include powdered gums such as agar, Karaya or tragacanth. Alginic acid and its sodium salt are also useful as disintegrants.

Binders may be used to hold the therapeutic  
10 agent together to form a hard tablet and include materials from natural products such as acacia, tragacanth, starch and gelatin. Others include methyl cellulose (MC), ethyl cellulose (EC) and carboxymethyl cellulose (CMC). Polyvinyl pyrrolidone (PVP) and  
15 hydroxypropylmethyl cellulose (HPMC) could both be used in alcoholic solutions to granulate the therapeutic.

An antifrictional agent may be included in the formulation of the therapeutic to prevent sticking during the formulation process. Lubricants may be used  
20 as a layer between the therapeutic and the die wall, and these can include but are not limited to; stearic acid including its magnesium and calcium salts, polytetrafluoroethylene (PTFE), liquid paraffin, vegetable oils and waxes. Soluble lubricants may also  
25 be used such as sodium lauryl sulfate, magnesium lauryl sulfate, polyethylene glycol of various molecular weights, Carbowax 4000 and 6000.

Glidants that might improve the flow properties of the drug during formulation and to aid  
30 rearrangement during compression might be added. The glidants may include starch, talc, pyrogenic silica and hydrated silicoaluminate.

To aid dissolution of the therapeutic into the aqueous environment a surfactant might be added as  
35 a wetting agent. Surfactants may include anionic detergents such as sodium lauryl sulfate, dioctyl

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sodium sulfosuccinate and dioctyl sodium sulfonate. Cationic detergents might be used and could include benzalkonium chloride or benzethonium chloride. The list of potential nonionic detergents that could be  
5 included in the formulation as surfactants are lauromacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 50 and 60, glycerol monostearate, polysorbate 40, 60, 65 and 80, sucrose fatty acid ester, methyl cellulose and  
10 carboxymethyl cellulose. These surfactants could be present in the formulation of the protein or derivative either alone or as a mixture in different ratios.

Additives which potentially enhance uptake of the protein (or derivative) are for instance the fatty  
15 acids oleic acid, linoleic acid and linolenic acid.

Controlled release formulation may be desirable. The drug could be incorporated into an inert matrix which permits release by either diffusion or leaching mechanisms e.g., gums. Slowly degenerating  
20 matrices may also be incorporated into the formulation, e.g., alginates, polysaccharides. Another form of a controlled release of this therapeutic is by a method based on the Oros therapeutic system (Alza Corp.), i.e., the drug is enclosed in a semipermeable membrane  
25 which allows water to enter and push drug out through a single small opening due to osmotic effects. Some enteric coatings also have a delayed release effect.

Other coatings may be used for the formulation. These include a variety of sugars which  
30 could be applied in a coating pan. The therapeutic agent could also be given in a film coated tablet and the materials used in this instance are divided into 2 groups. The first are the nonenteric materials and include methyl cellulose, ethyl cellulose, hydroxyethyl  
35 cellulose, methylhydroxy-ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl-methyl cellulose, sodium

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carboxy-methyl cellulose, providone and the polyethylene glycols. The second group consists of the enteric materials that are commonly esters of phthalic acid.

5           A mix of materials might be used to provide the optimum film coating. Film coating may be carried out in a pan coater or in a fluidized bed or by compression coating.

Also contemplated herein is pulmonary  
10 delivery of the present protein (or derivatives thereof). The protein (or derivative) is delivered to the lungs of a mammal while inhaling and traverses across the lung epithelial lining to the blood stream. (Other reports of this include Adjei et al.,  
15 Pharmaceutical Research 7: 565-569 (1990); Adjei et al., International Journal of Pharmaceutics 63: 135-144 (1990) (leuprolide acetate); Braquet et al., Journal of Cardiovascular Pharmacology 13 (suppl. 5): s.143-146  
20 Medicine 3: 206-212 (1989) ( $\alpha$ 1-antitrypsin); Smith et al., J. Clin. Invest. 84: 1145-1146 (1989) ( $\alpha$ 1-proteinase); Oswein et al., "Aerosolization of Proteins", Proceedings of Symposium on Respiratory Drug Delivery II, Keystone, Colorado, March, 1990  
25 (recombinant human growth hormone); Debs et al., The Journal of Immunology 140: 3482-3488 (1988) (interferon  $\gamma$  and tumor necrosis factor  $\alpha$ ) and U.S. Patent No. 5,284,656 (granulocyte colony stimulating factor).

Contemplated for use in the practice of this  
30 invention are a wide range of mechanical devices designed for pulmonary delivery of therapeutic products, including but not limited to nebulizers, metered dose inhalers, and powder inhalers, all of which are familiar to those skilled in the art.



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Some specific examples of commercially available devices suitable for the practice of this invention are the Ultravent nebulizer, manufactured by Mallinckrodt, Inc., St. Louis, Missouri; the Acorn II  
5 nebulizer, manufactured by Marquest Medical Products, Englewood, Colorado; the Ventolin metered dose inhaler, manufactured by Glaxo Inc., Research Triangle Park, North Carolina; and the Spinhaler powder inhaler, manufactured by Fisons Corp., Bedford, Massachusetts.

10 All such devices require the use of formulations suitable for the dispensing of protein (or analog or derivative). Typically, each formulation is specific to the type of device employed and may involve the use of an appropriate propellant material, in  
15 addition to diluents, adjuvants and/or carriers useful in therapy.

The protein (or derivative) should most advantageously be prepared in particulate form with an average particle size of less than 10  $\mu\text{m}$  (or microns),  
20 most preferably 0.5 to 5  $\mu\text{m}$ , for most effective delivery to the distal lung.

Carriers include carbohydrates such as trehalose, mannitol, xylitol, sucrose, lactose, and sorbitol. Other ingredients for use in formulations  
25 may include DPPC, DOPE, DSPC and DOPC. Natural or synthetic surfactants may be used. Polyethylene glycol may be used (even apart from its use in derivatizing the protein or analog). Dextrans, such as cyclodextran, may be used. Bile salts and other  
30 related enhancers may be used. Cellulose and cellulose derivatives may be used. Amino acids may be used, such as use in a buffer formulation.

Also, the use of liposomes, microcapsules or microspheres, inclusion complexes, or other types of  
35 carriers is contemplated.

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Nasal delivery of the protein (or analog or derivative) is also contemplated. Nasal delivery allows the passage of the protein to the blood stream directly after administering the therapeutic product to the nose, without the necessity for deposition of the product in the lung. Formulations for nasal delivery include those with dextran or cyclodextran. Delivery via transport across other mucus membranes is also contemplated.

#### Dosage

One skilled in the art will be able to ascertain effective dosages by administration and observing the desired therapeutic effect. Due to the modification of an AGP-1 protein by fusion to an Fc, the present invention provides unexpected protein protection from degradation, and increases circulation time and stability, when compared to a soluble AGP-1 protein. One skilled in the art, therefore, will be able to ascertain from these changes that an effective dosage may require lower doses or less frequent dosing.

Preferably, the formulation of the molecule will be such that between about .10 µg/kg and 10 mg/kg will yield the desired therapeutic effect. The frequency of administration may be readily determined by one skilled in the art. The effective dosages may be determined using diagnostic tools over time. For example, a diagnostic for measuring the amount of AGP-1 protein or Fc-AGP-1 fusion protein in the blood (or plasma or serum) may first be used to determine endogenous levels of protein. Such diagnostic tools may be in the form of an antibody assay, such as an antibody sandwich assay. The amount of endogenous AGP-1 protein is quantified initially, and a baseline is determined. The therapeutic dosages are determined as the quantification of endogenous and exogenous AGP-1

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protein or Fc-AGP-1 fusion protein (that is, protein, variant, fragment or derivative found within the body, either self-produced or administered) is continued over the course of therapy. The dosages may therefore vary  
5 over the course of therapy, with a relatively high dosage being used initially, until therapeutic benefit is seen, and lower dosages used to maintain the therapeutic benefits.

10           The following examples are offered to more fully illustrate the invention, but are not construed as limiting the scope thereof.

15

## EXAMPLE 1

## Production of AGP-1 Fusion Proteins

The fusion proteins Fc-huAGP-1 (95-281), Fc-huAGP-1 (114-281), Fc-muAGP-1 (99-291) and Fc-muAGP-1  
20 (120-291) were constructed by the following procedures.

The human IgG<sub>1</sub> Fc region was PCR amplified with the following set of oligonucleotide primers:

5' TCT CCA AGC TTG AGC CCA AAT CTT GTG ACA AAA C 3'  
25 (SEQ ID NO. 1)  
5' TCT CCC TTA AGT TTA CCC GGA GAC AGG GAG AG 3' (SEQ ID NO. 2)

PCR reaction was carried out in a volume of 50  $\mu$ l with  
30 1 unit of vent DNA polymerase (New England Biolabs) in 20 mM Tris-HCl pH8.8, 10 mM KCl, 10 mM  $(\text{NH}_4)_2\text{SO}_4$ , 0.1% Triton-X100, 10  $\mu$ M of each dNTP, 1 $\mu$ M of each primer and 10 ng of RANKFc/pCEPP4 template (Hsu et al, Proc. Natl. Acad. Sci. USA 96, 3540-3545 (1999)). Reactions were  
35 performed in 94°C for 30 sec., 55°C for 30 sec., and

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72°C for 1 min, for a total of 16 cycles. The PCR fragment was isolated by electrophoresis through 1% agarose and purification by the Geneclean procedure (Bio 101, Inc.). The PCR fragment creates a HindIII restriction site at 5' end and a AflIII restriction site at 3' end. The HindIII-AflIII digested PCR fragment was then subcloned into the pCEP4 vector (Invitrogen) to create Fc/pCEP4.

A murine OPG signal peptide having the following amino acid sequence:

MNKWLCCALLVLLDIIEWTTQ (SEQ ID NO. 3)

was created by annealing the following set of oligonucleotides:

5' CTA GCA CCA TGA ACA AGT GGC TGT GCT GCG CAC TCC  
TGG TGC TCC TGG ACA TCA TTG AAT GGA CAA CCC AGA 3' (SEQ  
ID NO. 4)

5' AGC TTC TGG GTT GTC CAT TCA ATG ATG TCC AGG AGC  
ACC AGG AGT GCG CAG CAC AGC CAC TTG TTC ATG GTG 3' (SEQ  
ID NO. 5)

The annealing was carried out by heating 10  $\mu$ M of each oligonucleotide in a total volume of 20  $\mu$ l at 94°C for 5 minutes, and then cooling gradually to room temperature. The annealed oligonucleotides, creating NheI restriction site overhang at 5' end and HindIII restriction site overhang at 3' end, were cloned into Fc-pCEP4 vector inframe N-terminal to the human IgG1 Fc region to generate SO-Fc/pCEP4 vector. A linker which encodes two irrelevant amino acids (QK) was introduced between the OPG signal peptide and human IgG1 Fc. An extra irrelevant amino acid (K) was introduced by the HindIII site between the OPG signal peptide and human IgG1 Fc.

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The baculovirus expression system chosen to express nucleic acid sequences encoding Fc-AGP-1 fusion proteins is based on the BAC-TO-BACK expression system  
5 (Life Technologies, Gaithersburg, MD) with the following modifications.

The major capsid protein promoter sequence (GenBank Acc. No. M22978) was PCR amplified from the  
10 Bac-N-Blue™ linear AcMNPV DNA purchased from Invitrogen (Carlsbad, CA) with the following primers:

5' ATT ATT GAT ATC GCA TGC TTG TTC GCC ATC GTG GAA TC  
(SEQ ID NO. 6)  
15 5' AAT CCG GAA TAT TGT TGC CGT TAT AAA TAT GGA C (SEQ  
ID NO. 7)

The N-terminal coding sequence of the first 12 codons of the polyhedrin gene with the methionine start codon  
20 mutated from ATG to ATT and the following MCS were PCR amplified from the vector pBlueBac4 DNA (Invitrogen) with the following primers:

5' AAC GGC AAC AAT ATT CCG GAT TAT TCA TAC CGT CC (SEQ  
25 ID NO. 8)  
5' ACT TCA AGG AGA ATT TCC (SEQ ID NO. 9)

The resulting two fragments share overlapping sequence and a second round of PCR was performed to fuse them  
30 together. The polyhedrin promoter and the multiple cloning sites (MCS) between the SnaBI and HindIII site present in pFastBac1 was then replaced with the EcoRV and HindIII digested fragment containing capsid promoter and the new MCS. The resulting vector is  
35 named pFC. To further modify the MCS, the following

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pair of primers were annealed together and ligated to the pFC vector digested with NheI and HindIII.

5' CTA GCT CTA GAC ATA TGG AAT TCC TGC AGC AGC TGG TAC  
5 CTC GAG GATCCA AGC TTG TCG ACT (SEQ ID NO. 10)  
5' AGC TAG TCG ACA AGC TTG GAT CCT CGA GGT ACC AGC TGC  
TGC AGG AAT TCC ATA TGT CTA GAG (SEQ ID NO. 11)

The resulting vector is named pFC1 and is used as the  
10 donor vector for all baculovirus expression work described here.

To generate Fc-AGP-1 sequences in a  
baculovirus expression vector, the SO-Fc/pCEP4 was  
15 first used as template for PCR amplification of OPG signal peptide followed by human IgGyl Fc region with the following primers:

5' GGG CGT GCT AGC CAC CAT GAA CAA GTG GCT GTG CTG C 3'  
20 (SEQ ID NO. 12)  
5' AGC TCC TTC TGC AGG TGG AAC AGC TGT TTA CCC GGA GAC  
AGG GAG 3' (SEQ ID NO. 13)

PCR reactions were carried out under conditions similar  
25 to those described above. The PCR fragment creates NheI restriction site at 5' end and PstI restriction site at 3' end, and was subsequently cloned into pFC1 vector to create SO-Fc/pFC1.

30 Human AGP-1(114-281) was PCR amplified from human AGP-1 and human IgGyl cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:

35 5' CTC CGG GTA AAG TGA GAG AAA GAG GTC CTC AG 3' (SEQ ID NO. 14)

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5' TTC TCT CAC TTT ACC CGG AGA CAG GGA G 3' (SEQ ID NO. 15)

5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 16)

5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 17)

5

Human AGP-1(95-281) was PCR amplified from human AGP-1 and human IgG $\gamma$ 1 cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:

10

5' CCG GGT AAA ACT TCT GAG GAA ACC ATT TCT AC 3' (SEQ ID NO. 18)

5' TCC TCA GAA GTT TTA CCC GGA GAC AGG GAG AG 3' (SEQ ID NO. 19)

15

5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 20)

5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 21)

Murine AGP-1(120-291) was PCR amplified from murine AGP-1 and human IgG  $\gamma$ 1 cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:

20

5' CCG GGT AAA GGT GGA AGA CCT CAG AAA GTG 3' (SEQ ID NO. 22)

25

5' GAG GTC TTC CAC CTT TAC CCG GAG ACA GGG AG 3' (SEQ ID NO. 23)

5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 24)

5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 25)

30

Murine AGP-1(99-291) was PCR amplified from human AGP-1 and human IgG  $\gamma$ 1 cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:

35

5' CTC CGG GTA AAA CCT TTC AGG ACA CCA TTT CTA C 3' (SEQ ID NO. 26)

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5' CCT GAA AGG TTT TAC CCG GAG ACA GGG AG 3' (SEQ ID NO. 27)

5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 28)

5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 29)

5

The PCR was carried out in similar conditions as described above. The resulting PCR fusion fragments were digested with NsiI and XhoI restriction enzymes and cloned in frame C-terminal to the human IgG1 Fc region in SO-Fc/pFC1 at NsiI and XhoI sites. The final pFC1 constructs expressing Fc-AGP-1 fusion proteins were subject to sequencing analysis.

Transformation of the donor plasmids, selection of the recombinant bacmids and isolation of the recombinant bacmid DNAs followed the protocols provided by the manufacturer (Life Technologies). The recombinant bacmid DNAs described above were transfected into insect SF-9 cells by standard calcium chloride precipitation method to generate recombinant viruses using Grace's insect cell medium with supplements (Invitrogen) and 10 % fetal bovine serum. The viruses were subsequently amplified using the same cell line and medium. The final titer of the viruses was estimated to be around  $10^7$  (pfu/ml) based on previous experiences. To produce Fc-AGP-1 fusion proteins, shake flask expression was set up to infect insect Hi-Five cells at a density of  $2 \times 10^6$  cells/ml grown in serum-free Ex-CELL405 medium from JRH Biosciences (Lenexa, KS). The amount of virus preparation used was about 1/10 of the total cell culture volume. Time course samples were collected and analyzed by Western blot analysis in a pilot experiment and the harvest time was determined to be at 50-55 hours post-infection. Some product degradation was observed at later hours and when more cells ( $> 10\%$ ) started to show signs of death. The harvested conditioned medium was filter-



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sterilized immediately and kept at -80°C until purification step.

5

## EXAMPLE 2

## Purification of AGP-1 Fusion Proteins

Fc-AGP-1 fusion proteins were purified using Pharmacia Protein A Sepharose. The resin was  
10 equilibrated with TBS containing 20mM Tris pH7.0 and 150mM NaCl before applying the media. Complete protease inhibitor cocktail (Boehringer-Mannheim) was added to the media according to the manufacturer's instructions. The media was loaded, the column washed  
15 with TBS, and protein was eluted using Gentle Elution buffer (Pierce, Rockford, IL). Protein containing fractions were pooled and submitted for in vitro analysis.

20

## EXAMPLE 3

## Production of AGP-1 Protein

PCR amplification employing the following  
25 primer pair and human AGP-1 cDNA template are used to generate various forms of human AGP-1.

5' ATT TGA TTC TAG AAG GAG GAA TAA CAT ATG GTT CGT GAA  
CGT GGT CCA CAG CGT GTA GCA 3' (SEQ ID NO. 30)

30

5' TAT CCG CGG ATC CTC GAG TTA GCC AAC TAA AAA GGC CCC  
GAA 3' (SEQ ID NO. 31)

One primer introduces unique XbaI and NdeI  
35 restrictions sites, an initiating Methionine codon, and optimized codons for the amino terminal protein of the

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gene. The other primer of the pair introduces a TAA stop codon, and a unique XhoI site following the carboxyl terminus of the gene. PCR and thermocycling was performed using standard recombinant DNA methodology. The PCR products were purified, restriction digested and inserted into the unique XbaI and XhoI sites of the vector pAMG21 (ATCC accession no. 98113), and transformed into the prototrophic E.coli strain 2596. The resulting construct pAMG21-huAGP-1 (114-281) was engineered to encode human AGP-1 from amino acids 114-281 (see Figure 2). After transformation, clones were selected, plasmid DNA was isolated and the sequence of the AGP-1 gene insert was confirmed.

Human AGP-1(114-281) was purified as follows. 40.5g of E. coli cell paste was homogenized in 20mM Tris, 10mM EDTA, pH7.5 with Complete protease inhibitors in a total volume of 250mL. Cells were lysed with two passes in a microfluidizer at 70psi and then centrifuged at 14,000 rpm in a JA14 rotor for 60min. The supernatant was filtered through 0.45µm and 0.22µm pore size filters. Aliquots were stored at -20°C. One 10mL aliquot was thawed and centrifuged. The supernatant was diluted 1:2 with 20mM Tris pH 7.5, adjusted to pH 7.5 then filtered through a 0.45µm filter. The sample was then applied to an 5mL SP Hi Trap column which had been equilibrated with the above buffer. The protein was eluted using a gradient from 0-0.5M NaCl in 20mM Tris pH 7.5. AGP-1 containing fractions were concentrated and then diafiltered into PBS pH6.2 using a centrprep 30. The concentrated fraction was estimated to be 90% pure by SDS-PAGE stained with Coomassie Blue.

## EXAMPLE 4

Biological Activity of AGP-1 and AGP-1 Fusion Proteins

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Jurkat cells (ATCC No. \_\_\_\_ ) were maintained in RPMI medium 1640 containing 10% fetal calf serum, 100 mg/ml penicillin G, and 100 mg/ml streptomycin (GIBCO). To study apoptotsis induced by FcAGP-1 recombina<sup>5</sup> nt protein, 250 ml of Jurkat cells (5 x 10<sup>5</sup> cells/ml) were seeded to each well of a 96 well plate. Cells were incubated in 5% CO<sub>2</sub> at 37°C with indicated concentrations of hu AGP-1 (114-281), huFcAGP-1 (114-281), or human IgG (Sigma) for 24 hours. Alamar Blue<sup>10</sup> (Biosource Inc.) was added to each well in an amount equal to 10% of the culture volume. Cells were incubated for another 8 hours. Fluorescence was measured with excitation wavelength at 530nm and emission wavelength at 590nm in Bio-TEK FL500<sup>15</sup> Fluorescence Plate Reader (Bio-TEK Instruments Inc.). Each experiment was performed in duplicates. The results shown in Figure 7 indicate enhanced apoptosis activity of FcAGP-1 compared to soluble AGP-1.

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\* \* \*

While the present invention has been described in terms of preferred embodiments, it is understood that variations and modifications will occur<sup>25</sup> to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent variations which come within the scope of the invention as claimed.

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## WHAT IS CLAIMED IS:

1. A protein having a formula selected from the group consisting of:  $R_1 - R_2$  and  $R_1 - L - R_2$ , wherein  
5  $R_1$  is a Fc protein, or variant or fragment thereof,  $R_2$  is an AGP-1 protein, or variant or fragment thereof, and L is a linker.
2. The protein according to claim 1, where  
10 in the Fc protein is selected from the group consisting of:
  - (a) the Fc amino acid sequences as set forth in Figure 1;
  - (b) the amino acid sequence of subpart (a)  
15 having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to Figure 1):
    - (i) one or more cysteine residues;
    - (ii) one or more tyrosine residues;
    - 20 (iii) cysteine at position 5 deleted or substituted with an alanine;
    - (iv) leucine at position 20 deleted or substituted with glutamine;
    - (v) glutamic acid at position 103  
25 deleted or substituted with an alanine;
    - (vi) lysine at position 105 deleted or substituted with an alanine;
    - (vii) lysine at position 107 deleted or substituted with an alanine;
    - 30 (viii) deletion or substitution of one or more of the amino acids at positions 1, 2, 3, 4, and 5;
    - (ix) one or more residues substituted or deleted to ablate the Fc receptor binding site;

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(x) one or more residues substituted or deleted to ablate the complement (C1q) binding site; and

(xi) a combination of subparts i-x;

5 (c) the amino acid sequence of subparts (a) or (b) having a methionyl residue at the N-terminus;

(d) the Fc protein, or variant, fragment or derivative thereof, of any of subparts (a) through  
10 (c) comprised of a chemical moiety connected to the protein moiety;

(e) a derivative of subpart (d) wherein said chemical moiety is a water soluble polymer moiety;

(f) a derivative of subpart (e) wherein said  
15 water soluble polymer moiety is polyethylene glycol; and

(g) a derivative of subpart (e) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

20

3. The protein according to claim 1, wherein the AGP-1 protein, or variant, fragment or derivative thereof, is selected from the group consisting of:

25 (a) the amino acid sequence X-281 wherein X is any residue from 95 to 114 inclusive as shown in Figure 2 (SEQ ID NO:34);

(b) the amino acid sequence of subpart (a) having a methionyl residue at the N-terminus.

30 (c) the AGP-1 protein, or variant, fragment or derivative thereof, of any of subparts (a) and (b) comprised of a chemical moiety connected to the protein moiety;

(d) a derivative of subpart (c) wherein said  
35 chemical moiety is a water soluble polymer moiety;

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(e) a derivative of subpart (d) wherein said water soluble polymer moiety is polyethylene glycol;

5 (f) A derivative of subpart (d) wherein said water soluble polymer moiety is a polyamino acid moiety; and

(g) a derivative of subpart (d) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

10

4. The protein of claim 1 wherein the linker sequence is one or more amino acids selected from the group consisting of: Glycine, Asparagine, Serine, Threonine and Alanine.

15

5. The protein of claim 1 wherein the linker is selected from the group consisting of:

- 20 (a) ala-ala-ala;  
(b) ala-ala-ala-ala;  
(c) ala-ala-ala-ala-ala;  
(d) gly-gly;  
(e) gly-gly-gly;  
(f) gly-gly-gly-gly-gly;  
(g) gly-gly-gly-gly-gly-gly-gly;  
25 (h) gly-pro-gly;  
(i) gly-gly-pro-gly-gly;  
(j) chemical moiety; and  
(k) any combination of subparts (a) through (j).

30

6. A fusion protein comprising a Fc protein, or variant, fragment or derivative thereof, fused to the N-terminus of an AGP-1 protein, or variant, fragment or derivative thereof.

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7. A nucleic acid sequence encoding for a protein having the formula selected from the group consisting of:  $R_1 - R_2$  and  $R_1 - L - R_2$ , wherein  $R_1$  is a Fc protein, or variant or fragment thereof,  $R_2$  is an AGP-1 protein, or variant or fragment thereof, and L is a linker.

8. The nucleic acid sequence according to claim 7 encoding for a protein comprising an Fc protein, variant, fragment or derivative portion selected from the group consisting of:

(a) the Fc amino acid sequences as set forth in Figure 1 (SEQ ID NO: 32);

(b) the amino acid sequence of subpart (a) having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to Figure 1):

- (i) one or more cysteine residues;
- (ii) one or more tyrosine residues;
- (iii) cysteine at position 5 deleted or substituted with an alanine;
- (iv) leucine at position 20 deleted or substituted with glutamine;
- (v) glutamic acid at position 103 deleted or substituted with an alanine;
- (vi) lysine at position 105 deleted or substituted with an alanine;
- (vii) lysine at position 107 deleted or substituted with an alanine;
- (viii) deletion or substitution of one or more of the amino acids at positions 1, 2, 3, 4, and 5;
- (ix) one or more residues substituted or deleted to ablate the Fc receptor binding site;

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(x) one or more residues substituted or deleted to ablate the complement (Clq) binding site; and

(xi) a combination of subparts i-x;

5 (c) the amino acid sequence of subparts (a) or (b) having a methionyl residue at the N-terminus;

(d) the Fc protein, or variant, fragment or derivative thereof, of any of subparts (a) through  
10 (c) comprised of a chemical moiety connected to the protein moiety;

(e) a derivative of subpart (d) wherein said chemical moiety is a water soluble polymer moiety;

(f) a derivative of subpart (e) wherein said  
15 water soluble polymer moiety is polyethylene glycol; and

(g) a derivative of subpart (e) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

20

9. The nucleic acid sequence according to claim 7 encoding for a protein comprising an AGP-1 protein, variant, fragment or derivative portion selected from the group consisting of:

25 (a) the amino acid sequence X-281 wherein X is any residue from 95 to 114 inclusive as shown in Figure 2 (SEQ ID NO: 34);

(b) the amino acid sequence of subpart (a) having a methionyl residue at the N-terminus.

30 (c) the AGP-1 protein, or variant, fragment or derivative thereof, of any of subparts (a) and (b) comprised of a chemical moiety connected to the protein moiety;

(d) a derivative of subpart (c) wherein said  
35 chemical moiety is a water soluble polymer moiety;



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(e) a derivative of subpart (d) wherein said water soluble polymer moiety is polyethylene glycol;

5 (f) A derivative of subpart (d) wherein said water soluble polymer moiety is a polyamino acid moiety; and

(g) a derivative of subpart (d) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

10

10. The nucleic acid sequence of claim 7 encoding for a protein with a linker sequence of one or more amino acids selected from the group consisting of: Gly, Asn, Ser, Thr and Ala.

15

11. The nucleic acid sequence of claim 7 encoding for a protein with a linker selected from the group consisting of:

- 20 (a) ala, ala, ala;  
(b) ala-ala-ala-ala;  
(c) ala-ala-ala-ala-ala;  
(d) gly-gly;  
(e) gly-gly-gly;  
(f) gly-gly-gly-gly-gly;  
25 (g) gly-gly-gly-gly-gly-gly-gly;  
(h) gly-pro-gly;  
(i) gly-gly-pro-gly-gly;  
(j) a chemical moiety; and  
(k) any combination of subparts (a)

30 through (j).

12. A nucleic acid sequence encoding for a fusion protein having a Fc protein, or variant, fragment or derivative thereof, fused to the N-terminus  
35 of an AGP-1 protein, or a variant, fragment or derivative thereof.

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13. A vector comprising a nucleic acid sequence according to any of Claims 7 to 12 inclusive.

5           14. A prokaryotic or eukaryotic host cell containing the vector of claim 13.

10           15. A process for producing a protein of claims 1 or 6 comprising the steps of culturing, under suitable conditions, the host cell of claim 14, and isolating the protein produced.

15           16. The process of claim 15 further comprising the step of purifying the protein produced.

20           17. A pharmaceutical composition comprising an effective amount of a protein according to claims 1 or 6, in a pharmaceutically acceptable diluent, adjuvant or carrier.

25           18. A method of inducing apoptosis in a tissue comprising administering a therapeutically effective amount of the protein according to Claim 1 or 6.

30           19. A method of treating of a disorder selected from the group consisting of proliferative disorder, an immune disorder or a viral-induced disorder comprising administering a therapeutically effective amount of the protein according to claims 1 or 6.

**FIGURE 1**  
**Amino acid sequence of hinge, CH2 and**  
**CH3 regions human IgGγ1**

Glu	Pro	Lys	Ser	Cys	Asp	Lys	Thr	His	Thr	Cys	Pro	Pro	Cys	Pro	Ala	1	5	10	15
Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro	20	25	30	
Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val	35	40	45	
Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	Tyr	Val	50	55	60	
Asp	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln	65	70	75	80
Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln	85	90	95	
Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala	100	105	110	
Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro	115	120	125	
Arg	Glu	Pro	Gln	Val	Tyr	Thr	Leu	Pro	Pro	Ser	Arg	Asp	Glu	Leu	Thr	130	135	140	
Lys	Asn	Gln	Val	Ser	Leu	Thr	Cys	Leu	Val	Lys	Gly	Phe	Tyr	Pro	Ser	145	150	155	160
Asp	Ile	Ala	Val	Glu	Trp	Glu	Ser	Asn	Gly	Gln	Pro	Glu	Asn	Asn	Tyr	165	170	175	
Lys	Thr	Thr	Pro	Pro	Val	Leu	Asp	Ser	Asp	Gly	Ser	Phe	Phe	Leu	Tyr	180	185	190	
Ser	Lys	Leu	Thr	Val	Asp	Lys	Ser	Arg	Trp	Gln	Gln	Gly	Asn	Val	Phe	195	200	205	
Ser	Cys	Ser	Val	Met	His	Glu	Ala	Leu	His	Asn	His	Tyr	Thr	Gln	Lys	210	215	220	
Ser	Leu	Ser	Leu	Ser	Pro	Gly	Lys									225	230		

**FIGURE 2**  
**Amino acid and nucleic acid sequence**  
**of Fc-huAGP-1 (95-281)**

GGCTGACTTA CAGCAGTCAG ACTCTGACAG GATC ATG GCT ATG ATG GAG GTC	52
Met Ala Met Met Glu Val	
1 5	
CAG GGG GGA CCC AGC CTG GGA CAG ACC TGC GTG CTG ATC GTG ATC TTC	100
Gln Gly Gly Pro Ser Leu Gly Gln Thr Cys Val Leu Ile Val Ile Phe	
10 15 20	
ACA GTG CTC CTG CAG TCT CTC TGT GTG GCT GTA ACT TAC GTG TAC TTT	148
Thr Val Leu Leu Gln Ser Leu Cys Val Ala Val Thr Tyr Val Tyr Phe	
25 30 35	
ACC AAC GAG CTG AAG CAG ATG CAG GAC AAG TAC TCC AAA AGT GGC ATT	196
Thr Asn Glu Leu Lys Gln Met Gln Asp Lys Tyr Ser Lys Ser Gly Ile	
40 45 50	
GCT TGT TTC TTA AAA GAA GAT GAC AGT TAT TGG GAC CCC AAT GAC GAA	244
Ala Cys Phe Leu Lys Glu Asp Asp Ser Tyr Trp Asp Pro Asn Asp Glu	
55 60 65 70	
GAG AGT ATG AAC AGC CCC TGC TGG CAA GTC AAG TGG CAA CTC CGT CAG	292
Glu Ser Met Asn Ser Pro Cys Trp Gln Val Lys Trp Gln Leu Arg Gln	
75 80 85	
CTC GTT AGA AAG ATG ATT TTG AGA ACC TCT GAG GAA ACC ATT TCT ACA	340
Leu Val Arg Lys Met Ile Leu Arg Thr Ser Glu Glu Thr Ile Ser Thr	
90 95 100	
GTT CAA GAA AAG CAA CAA AAT ATT TCT CCC CTA GTG AGA GAA AGA GGT	388
Val Gln Glu Lys Gln Gln Asn Ile Ser Pro Leu Val Arg Glu Arg Gly	
105 110 115	
CCT CAG AGA GTA GCA GCT CAC ATA ACT GGG ACC AGA GGA AGA AGC AAC	436
Pro Gln Arg Val Ala Ala His Ile Thr Gly Thr Arg Gly Arg Ser Asn	
120 125 130	
ACA TTG TCT TCT CCA AAC TCC AAG AAT GAA AAG GCT CTG GGC CGC AAA	484
Thr Leu Ser Ser Pro Asn Ser Lys Asn Glu Lys Ala Leu Gly Arg Lys	
135 140 145 150	
ATA AAC TCC TGG GAA TCA TCA AGG AGT GGG CAT TCA TTC CTG AGC AAC	532
Ile Asn Ser Trp Glu Ser Ser Arg Ser Gly His Ser Phe Leu Ser Asn	
155 160 165	
TTG CAC TTG AGG AAT GGT GAA CTG GTC ATC CAT GAA AAA GGG TTT TAC	580
Leu His Leu Arg Asn Gly Glu Leu Val Ile His Glu Lys Gly Phe Tyr	
170 175 180	
TAC ATC TAT TCC CAA ACA TAC TTT CGA TTT CAG GAG GAA ATA AAA GAA	628
Tyr Ile Tyr Ser Gln Thr Tyr Phe Arg Phe Gln Glu Glu Ile Lys Glu	
185 190 195	
AAC ACA AAG AAC GAC AAA CAA ATG GTC CAA TAT ATT TAC AAA TAC ACA	676
Asn Thr Lys Asn Asp Lys Gln Met Val Gln Tyr Ile Tyr Lys Tyr Thr	
200 205 210	
AGT TAT CCT GAC CCT ATA TTG TTG ATG AAA AGT GCT AGA AAT AGT TGT	724
Ser Tyr Pro Asp Pro Ile Leu Leu Met Lys Ser Ala Arg Asn Ser Cys	
215 220 225 230	
TGG TCT AAA GAT GCA GAA TAT GGA CTC TAT TCC ATC TAT CAA GGG GGA	772
Trp Ser Lys Asp Ala Glu Tyr Gly Leu Tyr Ser Ile Tyr Gln Gly Gly	
235 240 245	

## FIGURE 2 (con't)

ATA TTT GAG CTT AAG GAA AAT GAC AGA ATT TTT GTT TCT GTA ACA AAT	820
Ile Phe Glu Leu Lys Glu Asn Asp Arg Ile Phe Val Ser Val Thr Asn	
250 255 260	
GAG CAC TTG ATA GAC ATG GAC CAT GAA GCC AGT TTT TTC GGG GCC TTT	868
Glu His Leu Ile Asp Met Asp His Glu Ala Ser Phe Phe Gly Ala Phe	
265 270 275	
TTA GTT GGC TAA CTGACCTGGA AAGAAAAAGC AATAACCTCA AAGTGACTAT	920
Leu Val Gly *	
280	
TCAGTTTTCA GGATGATACA CTATGAAGAT GTTTCAAAAA ATCTGACCAA AACAAACAAA	980
CAGAAAACAG AAAACAAAAA AACCTCTATG CAATCTGAGT AGAGCAGCCA CAACCAAAT	1040
TGTATACAAC ACACCATGTA	1060

**FIGURE 3**  
**Fc-huAGP-1(95-281)**

```

      10              30              50
GCTAGccaccATGAACAAGTGGCTGTGCTGCGCACTCCTGGTGCCTGGACATCATTGA
      M N K W L C C A L L V L L D I I E
      70              90              110
ATGGACAACCCAGAAGCTTGAGCCCAATCTTGTGACAAACTCACACATGCCACCGTG
      W T T O K L E P K S C D K T H T C P P C
      130              150              170
CCCAGCACCTGAACCTCTGGGGGACCGTCAGTCTTCTCTTCCCCCAAAACCCAAGGA
      P A P E L L G G P S V F L F P P K P K D
      190              210              230
CACCCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGA
      T L M I S R T P E V T C V V V D V S H E
      250              270              290
AGACCCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGAC
      D P E V K F N W Y V D G V E V H N A K T
      310              330              350
AAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTTCAGCGTCTTACCGTCTCT
      K P R E E Q Y N S T Y R V V S V L T V L
      370              390              410
GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGAAGGTCTCCAACAAAGCCCTCCC
      H Q D W L N G K E Y K C K V S N K A L P
      430              450              470
AGCCCCCATCGAGAAAACCATCTTCCAAAGCCAAAGGGCAGCCCCGAGAACACAGGTGT
      A P I E K T I S K A K G Q P R E P Q V Y
      490              510              530
CACCCCTGCCCCCATCCCGGATGAGCTGACCAAGAACCAGGTGAGCCTGACCTGCCTGGT
      T L P P S R D E L T K N Q V S L T C L V
      550              570              590
CAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAA
      K G F Y P S D I A V E W E S N G Q P E N
      610              630              650
CAACTACAAGACCACGCCTCCCGTGTGGACTCCGACGGCTCCTTCTTCTCTACAGCAA
      N Y K T T P P V L D S D G S F F L Y S K
      670              690              710
GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCA
      L T V D K S R W Q Q G N V F S C S V M H
      730              750              770
TGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAACTTC
      E A L H N H Y T Q K S L S L S P G K I T S
      790              810              830
TGAGGAAACCATTTCTACAGTTCAAGAAAAGCAACAAAATATTTCTCCCTAGTGAGAGA
      E E T I S T V Q E K Q Q N I S P L V R E
      850              870              890
AAGAGGTCTCAGAGAGTAGCAGCTCACATAACTGGGACAGAGGAAGAAGCAACACATT
      R G P Q R V A A H I T G T R G R S N T L
      910              930              950
GTCTTCTCCAAACTCCAAGAAATGAAAAGGCTCTGGGCCGCAAAATAAACTCTGGGAATC
      S S P N S K N E K A L G R K I N S W E S

```

FIGURE 3 (con't)

```

          970                      990                      1010
ATCAAGGAGTGGGCATTTCCTGAGCAACTTGCACTTGAGGAATGGCGAACTGGTCAT
S R S G H S F L S N L H L R N G E L V I
    1030                1050                1070
CCATGAAAAAGGGTTTACTACATCTATTCCCAAACATACTTCGATTTTCAGGAGGAAAT
H E K G F Y Y I Y S Q T Y F R F Q E E I
    1090                1110                1130
AAAAGAAAACACAAAGAACGACAAACAAATGGTCCAATATATTACAAATACACAAGTTA
K E N T K N D K Q M V Q Y I Y K Y T S Y
    1150                1170                1190
TCCTGACCCTATATTGTTGATGAAAAGTGCTAGAAATAGTTGTTGGTCTAAAGATGCAGA
P D P I L L M K S A R N S C W S K D A E
    1210                1230                1250
ATATGGACTCTATTCCATCTATCAAGGGGGAATATTTGAGCTTAAGGAAAATGACAGAAT
Y G L Y S I Y Q G G I F E L K E N D R I
    1270                1290                1310
TTTGTTTCTGTAAACAAATGAGCACTTGATAGACATGGACCATGAAGCCAGTTTTTTCGG
F V S V T N E H L I D M D H E A S F F G
    1330
GGCCTTTTTAGTTGGCTAAActcgag
A F L V G] *

```

**FIGURE 4**  
**Fc-huAGP-1 (114-281)**

```

      10              30              50
GCTAGccaccATGAACAAGTGGCTGTGCTGCGCACTCCTGGTGCTCCTGGACATCATTGA
      M N K W L C C A L L V L L D I I E
      70              90              110
ATGGACAACCCAGAAGCTTGAGCCCAAATCTTGTGACAAAACCTCACACATGCCCCACCGTG
      W T T Q K L E P K S C D K T H T C P P C
      130              150              170
CCCAGCACCTGAACTCCTGGGGGACCGTCAGTCTTCTCTCCCCCAAAACCCAAGGA
      P A P E L L G G P S V F L F P P K P K D
      190              210              230
CACCCCTCATGATCTCCCGGACCCCTGAGGTACATGCGTGGTGGTGGACGTGAGCCACGA
      T L M I S R T P E V T C V V V D V S H E
      250              270              290
AGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGAC
      D P E V K F N W Y V D G V E V H N A K T
      310              330              350
AAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGCGTCCTCACCCTCCT
      K P R E E Q Y N S T Y R V V S V L T V L
      370              390              410
GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCCC
      H Q D W L N G K E Y K C K V S N K A L P
      430              450              470
AGCCCCCATCGAGAAAACCATCTCCAAGCCAAAGGGCAGCCCCGAGAAACACAGGTGTA
      A P I E K T I S K A K G Q P R E P Q V Y
      490              510              530
CACCCCTGCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCTGCCTGGT
      T L P P S R D E L T K N Q V S L T C L V
      550              570              590
CAAAGGCTTCTATCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAA
      K G F Y P S D I A V E W E S N G Q P E N
      610              630              650
CAACTACAAGACCACGCCTCCCGTGTGGACTCCGACGGCTCCTTCTCTCTACAGCAA
      N Y K T T P P V L D S D G S F F L Y S K
      670              690              710
GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCA
      L T V D K S R W Q Q G N V F S C S V M H
      730              750              770
TGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAAGTGAG
      E A L H N H Y T Q K S L S L S P G K [V R
      790              810              830
AGAAAGAGGTCTCTCAGAGAGTAGCAGCTCACATAACTGGGACCAGAGGAAGAAGCAACAC
      E R G P Q R V A A H I T G T R G R S N T
      850              870              890
ATTGTCTTCTCCAAACTCCAAGAATGAAAAGGCTCTGGGCCGCAAAATAAACTCCTGGGA
      L S S P N S K N E K A L G R K I N S W E
      910              930              950
ATCATCAAGGAGTGGGCATTTCCTTCTGAGCAACTTGCACTTGAGGAATGGCGAACTGGT
      S S R S G H S F L S N L H L R N G E L V

```



FIGURE 4 (con't)

```

          970                      990                      1010
CATCCATGAAAAAGGGTTTACTACATCTATTCCCAAACATACTTTCGATTTCAGGAGGA
I H E K G F Y Y I Y S Q T Y F R F Q E E
    1030                      1050                      1070

AATAAAAGAAAAACACAAAGAACGACAAACAAATGGTCCAATATATTACAAATACACAAG
I K E N T K N D K Q M V Q Y I Y K Y T S
    1090                      1110                      1130

TTATCCTGACCCCTATATTGTTGATGAAAAGTGCTAGAAATAGTTGTTGGTCTAAAGATGC
Y P D P I L L M K S A R N S C W S K D A
    1150                      1170                      1190

AGAATATGGACTCTATTCCATCTATCAAGGGGGAATATTGAGCTTAAGGAAAATGACAG
E Y G L Y S I Y Q G G I F E L K E N D R
    1210                      1230                      1250

AATTTTGTCTCTGTAACAAATGAGCACTTGATAGACATGGACCATGAAGCCAGTTTTTT
I F V S V T N E H L I D M D H E A S F F
    1270

CGGGGCCTTTTGTAGTTGGCTAAactcgag
G A F L V G] *

```

**FIGURE 5**  
**Fc- $\mu$ AGP-1 (99-291)**

```

      10              30              50
GCTAGccaccATGAACAAGTGGCTGTGCTGCGCACTCCTGGTGCCTGGACATCATTTGA
  M N K W L C C A L L V L L D I I E
      70              90             110
ATGGACAACCCAGAAGCTTGAGCCCAAATCTTGTGACAAAACCTACACATGCCCCACCGT
  W T T O K L E P K S C D K T H T C P P C
      130             150             170
CCCAGCACCTGAACCTCTGGGGGACCGTCAGTCTTCTCTTCCCCCAAACCCAAGGA
  P A P E L L G G P S V F L F P P K P K D
      190             210             230
CACCTCATGATCTCCCGGACCCCTGAGGTACATGCGTGGTGGTGACGTGAGCCACGA
  T L M I S R T P E V T C V V V D V S H E
      250             270             290
AGACCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGAC
  D P E V K F N W Y V D G V E V H N A K T
      310             330             350
AAAGCCCGGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGCGTCTCACCGTCCT
  K P R E E Q Y N S T Y R V V S V L T V L
      370             390             410
GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAGCCCTCCC
  H Q D W L N G K E Y K C K V S N K A L P
      430             450             470
AGCCCCCATCGAGAAAACCATCTCCAAGCCAAAGGGCAGCCCCGAGAACACAGGTGTA
  A P I E K T I S K A K G Q P R E P Q V Y
      490             510             530
CACCTGCCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCTGCCTGGT
  T L P P S R D E L T K N Q V S L T C L V
      550             570             590
CAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAA
  K G F Y P S D I A V E W E S N G Q P E N
      610             630             650
CAACTACAAGACCACGCCTCCCGTGCTGGACTCCGACGGCTCCTTCTTCTCTACAGCAA
  N Y K T T P P V L D S D G S F L Y S K
      670             690             710
GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCA
  L T V D K S R W Q Q G N V F S C S V M H
      730             750             770
TGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAAacctt
  E A L H N H Y T Q K S L S L S P G K [T F
      790             810             830
tcAGGACACCATTCTTACAGTTCAGAAAAGCAGCTAAGTACTCCTCCCTTGCCCAGAGG
  Q D T I S T V P E K Q L S T P P L P R G
      850             870             890
TGGAAGACCTCAGAAAGTGGCAGCTCACATTACTGGGATCACTCGGAGAAGCAACTCAGC
  G R P Q K V A A H I T G I T R R S N S A
      910             930             950
TTTAATTCCAATCTCCAAGGATGGAAAGACCTTAGGCCAGAAGATTGAATCCTGGGAGTC
  L I P I S K D G K T L G Q K I E S W E S

```

FIGURE 5 (con't)

```

          970              990              1010
CTCTCGGAAAGGGCATTCAATTTCTCAACCACGTGCTCTTTAGGAATGGAGAGCTGGTCAT
S R K G H S F L N H V L F R N G E L V I
    1030              1050              1070
CGAGCAGGAGGGCCTGTATTACATCTATTCCCAAACATACTTCCGATTTGAGGAAGCTGA
E Q E G L Y Y I Y S Q T Y F R F Q E A E
    1090              1110              1130
AGACGCTTCCAAGATGGTCTCAAAGGACAAGGTGAGAACCAAACAGCTGGTGCAGTACAT
D A S K M V S K D K V R T K Q L V Q Y I
    1150              1170              1190
CTACAAGTACACCAGCTATCCGGATCCCATAGTGCTCATGAAGAGCGCCAGAAACAGCTG
Y K Y T S Y P D P I V L M K S A R N S C
    1210              1230              1250
TTGGTCCAGAGATGCCGAGTACGGACTGTACTCCATCTATCAGGGAGGATTGTTTCGAGCT
W S R D A E Y G L Y S I Y Q G G L F E L
    1270              1290              1310
AAAAAAAAATGACAGGATTTTGTCTGTGACAAATGAACATTTGATGGACCTGGATCA
K K N D R I F V S V T N E H L M D L D Q
    1330              1350
AGAAGCCAGCTTCTTTGGAGCCTTTTAAATTAATAActaactcgag
E A S F F G A F L I N] *

```

**FIGURE 6**  
**Fc-muAGP-1(120-291)**

```

      10              30              50
GCTAGccaccATGAACAAGTGGCTGTGCTGCCGACTCCTGGTGTCTGGACATCATTGA
      M N K W L C C A L L V L L D I I E
      70              90              110
ATGGACAACCCAGAAGCTTGAGCCCAAATCTTGTGACAAAACCTCACACATGCCCCACCGTG
      W T T Q K L E P K S C D K T H T C P P C
      130              150              170
CCCAGCACCTGAACCTCTGGGGGGACCGTCAGTCTTCTCTTCCCCCAAAACCCAAGGA
      P A P E L L G G P S V F L F P P K P K D
      190              210              230
CACCCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGA
      T L M I S R T P E V T C V V V D V S H E
      250              270              290
AGACCCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAATGCCAAGAC
      D P E V K F N W Y V D G V E V H N A K T
      310              330              350
AAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTGGTCAGCGTCTCACCGTCTCT
      K P R E E Q Y N S T Y R V V S V L T V L
      370              390              410
GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAAGCCCTCCC
      H Q D W L N G K E Y K C K V S N K A L P
      430              450              470
AGCCCCCATCGAGAAAACCATCTCCAAGCCAAAGGGCAGCCCCGAGAACACAGGTGTA
      A P I E K T I S K A K G Q P R E P Q V Y
      490              510              530
CACCCCTGCCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCTGCCTGGT
      T L P P S R D E L T K N Q V S L T C L V
      550              570              590
CAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCCGAGAA
      K G F Y P S D I A V E W E S N G Q P E N
      610              630              650
CAACTACAAGACCACGCCTCCCGTGTGGACTCCGACGGCTCCTTCTTCTCTACAGCAA
      N Y K T T P P V L D S D G S F F L Y S K
      670              690              710
GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCA
      L T V D K S R W Q Q G N V F S C S V M H
      730              750              770
TGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAAGGTGG
      E A L H N H Y T Q K S L S L S P G K [G G
      790              810              830
AAGACCTCAGAAAGTGGCAGCTCACATTACTGGGATCACTCGGAGAAGCAACTCAGCTTT
      R P Q K V A A H I T G I T R R S N S A L
      850              870              890
AATTCCAATCTCCAAGGATGGAAAGACCTTAGGCCAGAAGATTGAATCCTGGGAGTCCCT
      I P I S K D G K T L G Q K I E S W E S S
      910              930              950
TCGGAAAGGGCATTCAATTTCTCAACCACGTGCTCTTTAGGAATGGAGAGCTGGTCATCGA
      R K G H S F L N H V L F R N G E L V I E

```

FIGURE 6 (con't)

```

          970                      990                      1010
GCAGGAGGGCCTGTATTACATCTATTCCCAAACATACTTCCGATTTCAGGAAGCTGAAGA
  Q  E  G  L  Y  Y  I  Y  S  Q  T  Y  F  R  F  Q  E  A  E  D
    1030                      1050                      1070

CGCTTCCAAGATGGTCTCAAAGGACAAGGTGAGAACCAAACAGCTGGTGCAGTACATCTA
  A  S  K  M  V  S  K  D  K  V  R  T  K  Q  L  V  Q  Y  I  Y
    1090                      1110                      1130

CAAGTACACCAGCTATCCGGATCCCATAGTGCTCATGAAGAGCGCCAGAAACAGCTGTTG
  K  Y  T  S  Y  P  D  P  I  V  L  M  K  S  A  R  N  S  C  W
    1150                      1170                      1190

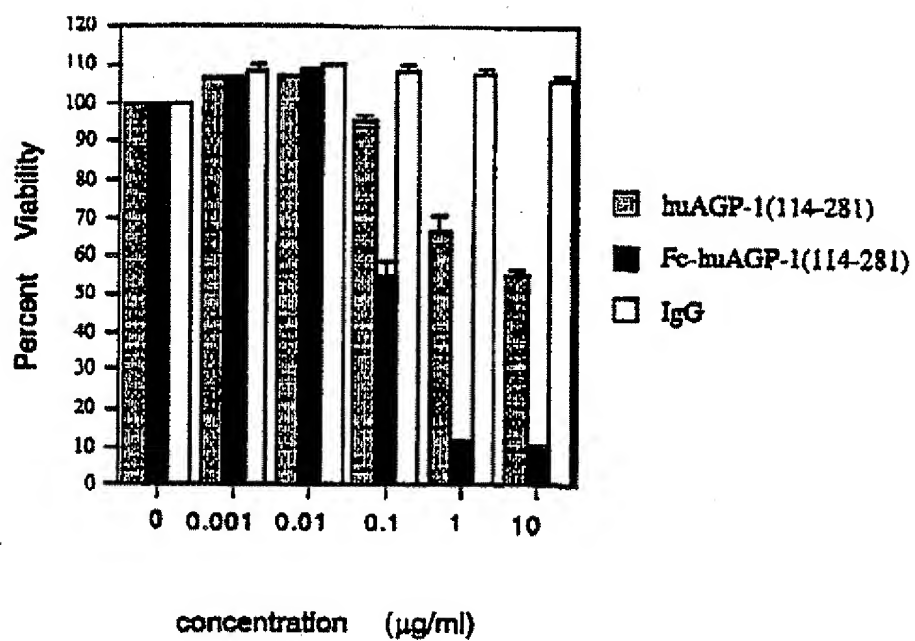
GTCCAGAGATGCCGAGTACGGACTGTACTCCATCTATCAGGGAGGATTGTTTCGAGCTAAA
  S  R  D  A  E  Y  G  L  Y  S  I  Y  Q  G  G  L  F  E  L  K
    1210                      1230                      1250

AAAAAATGACAGGATTTTGTTCCTGTGACAAATGAACATTTGATGGACCTGGATCAAGA
  K  N  D  R  I  F  V  S  V  T  N  E  H  L  M  D  L  D  Q  E
    1270                      1290

AGCCAGCTTCTTTGGAGCCTTTTAAATTAACATAactaactcgag
  A  S  F  F  G  A  F  L  I  N] *

```

FIGURE 7



## SEQUENCE LISTING

<110> Amgen Inc.

<120> AGP-1 Fusion Protein Compositions and Methods

<130> A-600

<140> A-600

<141> 1999-04-16

<160> 42

<170> PatentIn Ver. 2.0

<210> 1

<211> 33

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 1

tctccaagct tgagcccaaa tctgtgaca aaa

33

<210> 2

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 2

tctcccttaa gtttaccgg agacagggag ag

32

<210> 3

<211> 21

<212> PRT

<213> mouse

<400> 3

Met Asn Lys Trp Leu Cys Cys Ala Leu Leu Val Leu Leu Asp Ile Ile  
1 5 10 15

Glu Trp Thr Thr Gln  
20

<210> 4

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 4  
ctagcaccat gaacaagtgg ctgtgctgcg cactcctggg gctcctggac atcattgaat 60  
ggacaacca ga 72

<210> 5  
<211> 72  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 5  
agcttctggg ttgtccattc aatgatgtcc aggagcacca ggagtgcgca gcacagccac 60  
ttgttcatgg tg 72

<210> 6  
<211> 38  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 6  
attattgata tcgcatgctt gttcgccatc gtggaatc 38

<210> 7  
<211> 34  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 7  
aatccggaat attgttgccg ttataaatat ggac 34

<210> 8  
<211> 35  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 8  
aacggcaaca atattccgga ttattcatat ogtcc 35

<210> 9  
<211> 18  
<212> DNA  
<213> Artificial Sequence



&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 9

acttcaagga gaatttcc

18

&lt;210&gt; 10

&lt;211&gt; 63

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 10

ctagctctag acatatggaa ttctgcagc agctggtacc tcgaggatcc aagcttgctg 60

act

63

&lt;210&gt; 11

&lt;211&gt; 62

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 11

agctagtcga caagcttgga tcctcgaggt accagctgct gcaggaattc catatgtcta 60

gag

63

&lt;210&gt; 12

&lt;211&gt; 37

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 12

gggcgtgcta gccaccatga acaagtggct gtgctgc

37

&lt;210&gt; 13

&lt;211&gt; 45

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 13

agctccttct gcaggtggaa cagctgttta cccggagaca gggag

45

&lt;210&gt; 14

&lt;211&gt; 32

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 14

ctccgggtaa agtgagagaa agaggtcctc ag

32

<210> 15

<211> 28

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 15

ttctctcact ttaccgagac acagggag

28

<210> 16

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 16

cttcttctctc tacagcaagc

20

<210> 17

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 17

gttattgctc agcggtggca

20

<210> 18

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 18

ccgggtaaaa cttctgagga aaccatttct ac

32

<210> 19

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 19

tcctcagaag ttttaccgg agacagggag ag

32

<210> 20

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 20

cttcttcctc tacagcaagc

20

<210> 21

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 21

gttattgctc agcgggtggca

20

<210> 22

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 22

ccgggtaaag gtggaagacc tcagaaagtg

30

<210> 23

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: synthetic

<400> 23

gaggtcttcc acctttaccc ggagacaggg ag

32

<210> 24

<211> 20

<212> DNA

<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 24  
cttcttcctc tacagcaagc 20

<210> 25  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 25  
gttattgctc agcgggtggca 20

<210> 26  
<211> 34  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 26  
ctccgggtaa aacctttcag gacaccattt ctac 34

<210> 27  
<211> 29  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 27  
cctgaaaggt ttaccgcgga gacagggag 29

<210> 28  
<211> 20  
<212> DNA  
<213> Artificial Sequence

<220>  
<223> Description of Artificial Sequence: synthetic

<400> 28  
cttcttcctc tacagcaagc 20

<210> 29  
<211> 20  
<212> DNA  
<213> Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 29

gttattgctc agcggtggca

20

&lt;210&gt; 30

&lt;211&gt; 60

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 30

atttgattct agaaggagga ataacatatg gttcgtgaac gtggtccaca gcgtgtagca 60

&lt;210&gt; 31

&lt;211&gt; 42

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; Description of Artificial Sequence: synthetic

&lt;400&gt; 31

tatccgcgga tcctcgagtt agccaactaa aaaggccccc aa

42

&lt;210&gt; 32

&lt;211&gt; 232

&lt;212&gt; PRT

&lt;213&gt; Human

&lt;400&gt; 32

Glu	Pro	Lys	Ser	Cys	Asp	Lys	Thr	His	Thr	Cys	Pro	Pro	Cys	Pro	Ala
1				5					10					15	

Pro	Glu	Leu	Leu	Gly	Gly	Pro	Ser	Val	Phe	Leu	Phe	Pro	Pro	Lys	Pro
		20						25						30	

Lys	Asp	Thr	Leu	Met	Ile	Ser	Arg	Thr	Pro	Glu	Val	Thr	Cys	Val	Val
		35					40					45			

Val	Asp	Val	Ser	His	Glu	Asp	Pro	Glu	Val	Lys	Phe	Asn	Trp	Tyr	Val
		50				55					60				

Asp	Gly	Val	Glu	Val	His	Asn	Ala	Lys	Thr	Lys	Pro	Arg	Glu	Glu	Gln
65					70					75					80

Tyr	Asn	Ser	Thr	Tyr	Arg	Val	Val	Ser	Val	Leu	Thr	Val	Leu	His	Gln
			85						90					95	

Asp	Trp	Leu	Asn	Gly	Lys	Glu	Tyr	Lys	Cys	Lys	Val	Ser	Asn	Lys	Ala
		100						105					110		

Leu	Pro	Ala	Pro	Ile	Glu	Lys	Thr	Ile	Ser	Lys	Ala	Lys	Gly	Gln	Pro
		115					120						125		

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr  
 130 135 140

Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser  
 145 150 155 160

Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr  
 165 170 175

Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr  
 180 185 190

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe  
 195 200 205

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys  
 210 215 220

Ser Leu Ser Leu Ser Pro Gly Lys  
 225 230

&lt;210&gt; 33

&lt;211&gt; 1060

&lt;212&gt; DNA

&lt;213&gt; Human

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; Complement((35)..(877))

&lt;400&gt; 33

ggctgactta cagcagtcag actctgacag gata atg gct atg atg gag gtc cag 55  
 Met Ala Met Met Glu Val Gln  
 1 5

ggg gga ccc agc ctg gga cag acc tgc gtg ctg atc gtg atc ttc aca 103  
 Gly Gly Pro Ser Leu Gly Gln Thr Cys Val Leu Ile Val Ile Phe Thr  
 10 15 20

gtg ctc ctg cag tct ctc tgt gtg gct gta act tac gtg tac ttt acc 151  
 Val Leu Leu Gln Ser Leu Cys Val Ala Val Thr Tyr Val Tyr Phe Thr  
 25 30 35

aac gag ctg aag cag atg cag gac aag tac tcc aaa agt ggc att gct 199  
 Asn Glu Leu Lys Gln Met Gln Asp Lys Tyr Ser Lys Ser Gly Ile Ala  
 40 45 50 55

tgt ttc tta aaa gaa gat gac agt tat tgg gac ccc aat gac gaa gag 247  
 Cys Phe Leu Lys Glu Asp Asp Ser Tyr Trp Asp Pro Asn Asp Glu Glu  
 60 65 70

agt atg aac agc ccc tgc tgg caa gtc aag tgg caa ctc cgt cag ctc 295  
 Ser Met Asn Ser Pro Cys Trp Gln Val Lys Trp Gln Leu Arg Gln Leu  
 75 80 85

gtt aga aag atg att ttg aga acc tct gag gaa acc att tct aca gtt 343  
 Val Arg Lys Met Ile Leu Arg Thr Ser Glu Glu Thr Ile Ser Thr Val  
 90 95 100

caa gaa aag caa caa aat att tct ccc cta gtg aga gaa aga ggt cct 391

Gln Glu Lys Gln Gln Asn Ile Ser Pro Leu Val Arg Glu Arg Gly Pro  
 105 110 115  
 cag aga gta gca gct cac ata act ggg acc aga gga aga agc aac aca 439  
 Gln Arg Val Ala Ala His Ile Thr Gly Thr Arg Gly Arg Ser Asn Thr  
 120 125 130 135  
 ttg tct tct cca aac tcc aag aat gaa aag gct ctg ggc cgc aaa ata 487  
 Leu Ser Ser Pro Asn Ser Lys Asn Glu Lys Ala Leu Gly Arg Lys Ile  
 140 145 150  
 aac tcc tgg gaa tca tca agg agt ggg cat tca ttc ctg agc aac ttg 535  
 Asn Ser Trp Glu Ser Ser Arg Ser Gly His Ser Phe Leu Ser Asn Leu  
 155 160 165  
 cac ttg agg aat ggt gaa ctg gtc atc cat gaa aaa ggg ttt tac tac 583  
 His Leu Arg Asn Gly Glu Leu Val Ile His Glu Lys Gly Phe Tyr Tyr  
 170 175 180  
 atc tat tcc caa aca tac ttt cga ttt cag gag gaa ata aaa gaa aac 631  
 Ile Tyr Ser Gln Thr Tyr Phe Arg Phe Gln Glu Glu Ile Lys Glu Asn  
 185 190 195  
 aca aag aac gac aaa caa atg gtc caa tat att tac aaa tac aca agt 679  
 Thr Lys Asn Asp Lys Gln Met Val Gln Tyr Ile Tyr Lys Tyr Thr Ser  
 200 205 210 215  
 tat cct gac cct ata ttg ttg atg aaa agt gct aga aat agt tgt tgg 727  
 Tyr Pro Asp Pro Ile Leu Leu Met Lys Ser Ala Arg Asn Ser Cys Trp  
 220 225 230  
 tct aaa gat gca gaa tat gga ctc tat tcc atc tat caa ggg gga ata 775  
 Ser Lys Asp Ala Glu Tyr Gly Leu Tyr Ser Ile Tyr Gln Gly Gly Ile  
 235 240 245  
 ttt gag ctt aag gaa aat gac aga att ttt gtt tct gta aca aat gag 823  
 Phe Glu Leu Lys Glu Asn Asp Arg Ile Phe Val Ser Val Thr Asn Glu  
 250 255 260  
 cac ttg ata gac atg gac cat gaa gcc agt ttt ttc ggg gcc ttt tta 871  
 His Leu Ile Asp Met Asp His Glu Ala Ser Phe Phe Gly Ala Phe Leu  
 265 270 275  
 gtt ggc taactgacct ggaaagaaaa agcaataacc tcaaagtgac tattcagttt 927  
 Val Gly  
 280  
 tcaggatgat acactatgaa gatgtttcaa aaaatctgac caaaacaaac aaacagaaaa 987  
 cagaaaacaa aaaaacctct atgcaatctg agtagagcag ccacaaccaa aattgtatac 1047  
 aacacaccat gta 1060

<210> 34  
 <211> 281  
 <212> PRT  
 <213> Human

<400> 34  
 Met Ala Met Met Glu Val Gln Gly Gly Pro Ser Leu Gly Gln Thr Cys

1	5	10	15
Val Leu Ile	Val Ile Phe Thr	Val Leu Leu Gln Ser Leu	Cys Val Ala
20		25	30
Val Thr Tyr	Val Tyr Phe Thr	Asn Glu Leu Lys Gln Met	Gln Asp Lys
35		40	45
Tyr Ser Lys	Ser Gly Ile Ala Cys	Phe Leu Lys Glu Asp	Asp Ser Tyr
50		55	60
Trp Asp Pro	Asn Asp Glu Glu Ser	Met Asn Ser Pro Cys	Trp Gln Val
65		70	75
Lys Trp Gln	Leu Arg Gln Leu Val	Arg Lys Met Ile Leu	Arg Thr Ser
	85	90	95
Glu Glu Thr	Ile Ser Thr Val Gln	Glu Lys Gln Gln Asn	Ile Ser Pro
	100	105	110
Leu Val Arg	Glu Arg Gly Pro Gln	Arg Val Ala Ala His	Ile Thr Gly
	115	120	125
Thr Arg Gly	Arg Ser Asn Thr Leu	Ser Ser Pro Asn Ser	Lys Asn Glu
	130	135	140
Lys Ala Leu	Gly Arg Lys Ile Asn	Ser Trp Glu Ser Ser	Arg Ser Gly
145		150	155
His Ser Phe	Leu Ser Asn Leu His	Leu Arg Asn Gly Glu	Leu Val Ile
	165	170	175
His Glu Lys	Gly Phe Tyr Tyr Ile	Tyr Ser Gln Thr Tyr	Phe Arg Phe
	180	185	190
Gln Glu Glu	Ile Lys Glu Asn Thr	Lys Asn Asp Lys Gln	Met Val Gln
	195	200	205
Tyr Ile Tyr	Lys Tyr Thr Ser Tyr	Pro Asp Pro Ile Leu	Leu Met Lys
	210	215	220
Ser Ala Arg	Asn Ser Cys Trp Ser	Lys Asp Ala Glu Tyr	Gly Leu Tyr
225		230	235
Ser Ile Tyr	Gln Gly Gly Ile Phe	Glu Leu Lys Glu Asn	Asp Arg Ile
	245	250	255
Phe Val Ser	Val Thr Asn Glu His	Leu Ile Asp Met Asp	His Glu Ala
	260	265	270
Ser Phe Phe	Gly Ala Phe Leu Val	Gly	
	275	280	

<210> 35  
 <211> 1345  
 <212> DNA  
 <213> mouse

<220>  
 <221> CDS



&lt;222&gt; Complement((11)..(1333))

&lt;400&gt; 35

gctagccacc atg aac aag tgg ctg tgc tgc gca ctc ctg gtg ctc ctg	49
Met Asn Lys Trp Leu Cys Cys Ala Leu Leu Val Leu Leu	
1 5 10	
gac atc att gaa tgg aca acc cag aag ctt gag ccc aaa tct tgt gac	97
Asp Ile Ile Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp	
15 20 25	
aaa act cac aca tgc cca ccg tgc cca gca cct gaa ctc ctg ggg gga	145
Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly	
30 35 40 45	
ccg tca gtc ttc ctc ttc ccc cca aaa ccc aag gac acc ctc atg atc	193
Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile	
50 55 60	
tcc cgg acc cct gag gtc aca tgc gtg gtg gtg gac gtg agc cac gaa	241
Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu	
65 70 75	
gac cct gag gtc aag ttc aac tgg tac gtg gac ggc gtg gag gtg cat	289
Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His	
80 85 90	
aat gcc aag aca aag ccg cgg gag gag cag tac aac agc acg tac cgt	337
Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg	
95 100 105	
gtg gtc agc gtc ctc acc gtc ctg cac cag gac tgg ctg aat ggc aag	385
Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys	
110 115 120 125	
gag tac aag tgc aag gtc tcc aac aaa gcc ctc cca gcc ccc atc gag	433
Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu	
130 135 140	
aaa acc atc tcc aaa gcc aaa ggg cag ccc cga gaa cca cag gtg tac	481
Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr	
145 150 155	
acc ctg ccc cca tcc ccg gat gag ctg acc aag aac cag gtc agc ctg	529
Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu	
160 165 170	
acc tgc ctg gtc aaa ggc ttc tat ccc agc gac atc gcc gtg gag tgg	577
Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp	
175 180 185	
gag agc aat ggg cag ccg gag aac aac tac aag acc acg cct ccc gtg	625
Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val	
190 195 200 205	
ctg gac tcc gac ggc tcc ttc ttc ctc tac agc aag ctc acc gtg gac	673
Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp	
210 215 220	
aag agc agg tgg cag cag ggg aac gtc ttc tca tgc tcc gtg atg cat	721
Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His	

	225	230	235	
	gag gct ctg cac aac cac tac acg	cag aag agc ctc tcc ctg tct ccg		769
	Glu Ala Leu His Asn His Tyr Thr	Gln Lys Ser Leu Ser Leu Ser Pro		
	240	245	250	
	ggt aaa act tct gag gaa acc att tct aca gtt caa gaa aag caa caa			817
	Gly Lys Thr Ser Glu Glu Thr Ile Ser Thr Val Gln Glu Lys Gln Gln			
	255	260	265	
	aat att tct ccc cta gtg aga gaa aga ggt cct cag aga gta gca gct			865
	Asn Ile Ser Pro Leu Val Arg Glu Arg Gly Pro Gln Arg Val Ala Ala			
	270	275	280	285
	cac ata act ggg acc aga gga aga agc aac aca ttg tct tct cca aac			913
	His Ile Thr Gly Thr Arg Gly Arg Ser Asn Thr Leu Ser Ser Pro Asn			
	290	295	300	
	tcc aag aat gaa aag gct ctg ggc cgc aaa ata aac tcc tgg gaa tca			961
	Ser Lys Asn Glu Lys Ala Leu Gly Arg Lys Ile Asn Ser Trp Glu Ser			
	305	310	315	
	tca agg agt ggg cat tca ttc ctg agc aac ttg cac ttg agg aat ggc			1009
	Ser Arg Ser Gly His Ser Phe Leu Ser Asn Leu His Leu Arg Asn Gly			
	320	325	330	
	gaa ctg gtc atc cat gaa aaa ggg ttt tac tac atc tat tcc caa aca			1057
	Glu Leu Val Ile His Glu Lys Gly Phe Tyr Tyr Ile Tyr Ser Gln Thr			
	335	340	345	
	tac ttt cga ttt cag gag gaa ata aaa gaa aac aca aag aac gac aaa			1105
	Tyr Phe Arg Phe Gln Glu Glu Ile Lys Glu Asn Thr Lys Asn Asp Lys			
	350	355	360	365
	caa atg gtc caa tat att tac aaa tac aca agt tat cct gac cct ata			1153
	Gln Met Val Gln Tyr Ile Tyr Lys Tyr Thr Ser Tyr Pro Asp Pro Ile			
	370	375	380	
	ttg ttg atg aaa agt gct aga aat agt tgt tgg tct aaa gat gca gaa			1201
	Leu Leu Met Lys Ser Ala Arg Asn Ser Cys Trp Ser Lys Asp Ala Glu			
	385	390	395	
	tat gga ctc tat tcc atc tat caa ggg gga ata ttt gag ctt aag gaa			1249
	Tyr Gly Leu Tyr Ser Ile Tyr Gln Gly Gly Ile Phe Glu Leu Lys Glu			
	400	405	410	
	aat gac aga att ttt gtt tct gta aca aat gag cac ttg ata gac atg			1297
	Asn Asp Arg Ile Phe Val Ser Val Thr Asn Glu His Leu Ile Asp Met			
	415	420	425	
	gac cat gaa gcc agt ttt ttc ggg gcc ttt tta gtt ggctaactcg ag			1345
	Asp His Glu Ala Ser Phe Phe Gly Ala Phe Leu Val			
	430	435	440	

<210> 36  
 <211> 441  
 <212> PRT  
 <213> mouse

<400> 36

Met Asn Lys Trp Leu Cys Cys Ala Leu Leu Val Leu Leu Asp Ile Ile  
 1 5 10 15  
 Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp Lys Thr His  
 20 25 30  
 Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val  
 35 40 45  
 Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr  
 50 55 60  
 Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu  
 65 70 75 80  
 Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys  
 85 90 95  
 Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser  
 100 105 110  
 Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys  
 115 120 125  
 Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile  
 130 135 140  
 Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro  
 145 150 155 160  
 Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu  
 165 170 175  
 Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn  
 180 185 190  
 Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser  
 195 200 205  
 Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg  
 210 215 220  
 Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu  
 225 230 235 240  
 His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys Thr  
 245 250 255  
 Ser Glu Glu Thr Ile Ser Thr Val Gln Glu Lys Gln Gln Asn Ile Ser  
 260 265 270  
 Pro Leu Val Arg Glu Arg Gly Pro Gln Arg Val Ala Ala His Ile Thr  
 275 280 285  
 Gly Thr Arg Gly Arg Ser Asn Thr Leu Ser Ser Pro Asn Ser Lys Asn  
 290 295 300  
 Glu Lys Ala Leu Gly Arg Lys Ile Asn Ser Trp Glu Ser Ser Arg Ser  
 305 310 315 320  
 Gly His Ser Phe Leu Ser Asn Leu His Leu Arg Asn Gly Glu Leu Val

	325		330		335
Ile His Glu Lys Gly Phe Tyr Tyr	340	Ile Tyr Ser Gln Thr Tyr Phe Arg	345	350	
Phe Gln Glu Glu Ile Lys Glu Asn Thr Lys Asn Asp Lys Gln Met Val	355	360	365		
Gln Tyr Ile Tyr Lys Tyr Thr Ser Tyr Pro Asp Pro Ile Leu Leu Met	370	375	380		
Lys Ser Ala Arg Asn Ser Cys Trp Ser Lys Asp Ala Glu Tyr Gly Leu	385	390	395	400	
Tyr Ser Ile Tyr Gln Gly Gly Ile Phe Glu Leu Lys Glu Asn Asp Arg	405	410	415		
Ile Phe Val Ser Val Thr Asn Glu His Leu Ile Asp Met Asp His Glu	420	425	430		
Ala Ser Phe Phe Gly Ala Phe Leu Val	435	440			
<210> 37					
<211> 1288					
<212> DNA					
<213> mouse					
<220>					
<221> CDS					
<222> Complement((11)..(1279))					
<400> 37					
gctagccacc atg aac aag tgg ctg tgc tgc gca ctc ctg gtg ctc ctg					49
Met Asn Lys Trp Leu Cys Cys Ala Leu Leu Val Leu Leu					
1 5 10					
gac atc att gaa tgg aca acc cag aag ctt gag ccc aaa tct tgt gac					97
Asp Ile Ile Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp					
15 20 25					
aaa act cac aca tgc cca ccg tgc cca gca cct gaa ctc ctg ggg gga					145
Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly					
30 35 40 45					
ccg tca gtc ttc ctc ttc ccc cca aaa ccc aag gac acc ctc atg atc					193
Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile					
50 55 60					
tcc cgg acc cct gag gtc aca tgc gtg gtg gtg gac gtg agc cac gaa					241
Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu					
65 70 75					
gac cct gag gtc aag ttc aac tgg tac gtg gac ggc gtg gag gtg cat					289
Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His					
80 85 90					
aat gcc aag aca aag ccg cgg gag gag cag tac aac agc acg tac cgt					337
Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg					
95 100 105					

gtg gtc agc gtc ctc acc gtc ctg cac cag gac tgg ctg aat ggc aag	385
Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys	
110 115 120 125	
gag tac aag tgc aag gtc tcc aac aaa gcc ctc cca gcc ccc atc gag	433
Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu	
130 135 140	
aaa acc atc tcc aaa gcc aaa ggg cag ccc cga gaa cca cag gtg tac	481
Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr	
145 150 155	
acc ctg ccc cca tcc cgg gat gag ctg acc aag aac cag gtc agc ctg	529
Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu	
160 165 170	
acc tgc ctg gtc aaa ggc ttc tat ccc agc gac atc gcc gtg gag tgg	577
Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp	
175 180 185	
gag agc aat ggg cag ccg gag aac aac tac aag acc acg cct ccc gtg	625
Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val	
190 195 200 205	
ctg gac tcc gac ggc tcc ttc ttc ctc tac agc aag ctc acc gtg gac	673
Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp	
210 215 220	
aag agc agg tgg cag cag ggg aac gtc ttc tca tgc tcc gtg atg cat	721
Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His	
225 230 235	
gag gct ctg cac aac cac tac acg cag aag agc ctc tcc ctg tct ccg	769
Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro	
240 245 250	
ggg aaa gtg aga gaa aga ggt cct cag aga gta gca gct cac ata act	817
Gly Lys Val Arg Glu Arg Gly Pro Gln Arg Val Ala Ala His Ile Thr	
255 260 265	
ggg acc aga gga aga agc aac aca ttg tct tct cca aac tcc aag aat	865
Gly Thr Arg Gly Arg Ser Asn Thr Leu Ser Ser Pro Asn Ser Lys Asn	
270 275 280 285	
gaa aag gct ctg ggc cgc aaa ata aac tcc tgg gaa tca tca agg agt	913
Glu Lys Ala Leu Gly Arg Lys Ile Asn Ser Trp Glu Ser Ser Arg Ser	
290 295 300	
ggg cat tca ttc ctg agc aac ttg cac ttg agg aat ggc gaa ctg gtc	961
Gly His Ser Phe Leu Ser Asn Leu His Leu Arg Asn Gly Glu Leu Val	
305 310 315	
atc cat gaa aaa ggg ttt tac tac atc tat tcc caa aca tac ttt cga	1009
Ile His Glu Lys Gly Phe Tyr Tyr Ile Tyr Ser Gln Thr Tyr Phe Arg	
320 325 330	
ttt cag gag gaa ata aaa gaa aac aca aag aac gac aaa caa atg gtc	1057
Phe Gln Glu Glu Ile Lys Glu Asn Thr Lys Asn Asp Lys Gln Met Val	
335 340 345	

caa tat att tac aaa tac aca agt tat cct gac cct ata ttg ttg atg 1105  
 Gln Tyr Ile Tyr Lys Tyr Thr Ser Tyr Pro Asp Pro Ile Leu Leu Met  
 350 355 360 365

aaa agt gct aga aat agt tgt tgg tct aaa gat gca gaa tat gga ctc 1153  
 Lys Ser Ala Arg Asn Ser Cys Trp Ser Lys Asp Ala Glu Tyr Gly Leu  
 370 375 380

tat tcc atc tat caa ggg gga ata ttt gag ctt aag gaa aat gac aga 1201  
 Tyr Ser Ile Tyr Gln Gly Gly Ile Phe Glu Leu Lys Glu Asn Asp Arg  
 385 390 395

att ttt gtt tct gta aca aat gag cac ttg ata gac atg gac cat gaa 1249  
 Ile Phe Val Ser Val Thr Asn Glu His Leu Ile Asp Met Asp His Glu  
 400 405 410

gcc agt ttt ttc ggg gcc ttt tta gtt ggc taactcgag 1288  
 Ala Ser Phe Phe Gly Ala Phe Leu Val Gly  
 415 420

<210> 38

<211> 423

<212> PRT

<213> mouse

<400> 38

Met Asn Lys Trp Leu Cys Cys Ala Leu Leu Val Leu Leu Asp Ile Ile  
 1 5 10 15

Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp Lys Thr His  
 20 25 30

Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val  
 35 40 45

Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr  
 50 55 60

Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu  
 65 70 75 80

Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys  
 85 90 95

Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser  
 100 105 110

Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys  
 115 120 125

Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile  
 130 135 140

Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro  
 145 150 155 160

Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu  
 165 170 175

Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn

<400> 39

gctagccacc atg aac aag tgg ctg tgc tgc gca ctc ctg gtg ctc ctg	49
Met Asn Lys Trp Leu Cys Cys Ala Leu Val Leu Leu	
1 5 10	
gac atc att gaa tgg aca acc cag aag ctt gag ccc aaa tct tgt gac	97
Asp Ile Ile Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp	
15 20 25	
aaa act cac aca tgc cca ccg tgc cca gca oct gaa ctc ctg ggg gga	145
Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly	
30 35 40 45	
ccg tca gtc ttc ctc ttc ccc cca aaa ccc aag gac acc ctc atg atc	193
Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile	
50 55 60	
tcc cgg acc cct gag gtc aca tgc gtg gtg gtg gac gtg agc cac gaa	241
Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu	
65 70 75	
gac cct gag gtc aag ttc aac tgg tac gtg gac ggc gtg gag gtg cat	289
Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His	
80 85 90	
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Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr	
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Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu	
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cac att act ggg atc act cgg aga agc aac tca gct tta att cca atc    913
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 420 425

# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/US 00/08004

<b>A. CLASSIFICATION OF SUBJECT MATTER</b>		
IPC 7	C07K19/00 A61P37/00	C07K14/705 A61P31/12
C12N15/86	C12N5/10	A61K38/18
According to International Patent Classification (IPC) or to both national classification and IPC		
<b>B. FIELDS SEARCHED</b>		
Minimum documentation searched (classification system followed by classification symbols) IPC 7 C07K		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched		
Electronic data base consulted during the international search (name of data base and, where practical, search terms used) STRAND, MEDLINE, CANCERLIT, AIDSLINE, LIFESCIENCES, CHEM ABS Data, WPI Data, EPO-Internal, PAJ		
<b>C. DOCUMENTS CONSIDERED TO BE RELEVANT</b>		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 97 01633 A (IMMUNEX CORP) 16 January 1997 (1997-01-16) page 9, line 11 -page 13, line 8 page 21, line 5 -page 24, line 2 examples 8,9,11 claims	1-19
X	WO 97 46686 A (AMGEN INC) 11 December 1997 (1997-12-11) cited in the application page 3, line 13-24 claims 14-19 page 10, line 2-31	1,3,6,7, 9,12-17, 19
-/-		
<input checked="" type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex.		
* Special categories of cited documents : "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "&" document member of the same patent family		
Date of the actual completion of the international search  5 July 2000		Date of mailing of the international search report  27/07/2000
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016		Authorized officer  Covone, M

# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/08004

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>SHERIDAN J P ET AL: "Control of TRAIL-induced apoptosis by a family of signaling and decoy receptors" SCIENCE,US,AMERICAN ASSOCIATION FOR THE ADVANCEMENT OF SCIENCE,, vol. 277, 8 August 1997 (1997-08-08), pages 818-821, XP002065023 ISSN: 0036-8075 the whole document</p>	1-19
A	<p>KIM J -K ET AL: "IDENTIFYING AMINO ACID RESIDUES THAT INFLUENCE PLASMA CLEARANCE OF MURINE IGG1 FRAGMENTS BY SITE-DIRECTED MUTAGENESIS" EUROPEAN JOURNAL OF IMMUNOLOGY,DE,WEINHEIM, vol. 24, no. 3, 1 January 1994 (1994-01-01), pages 542-548, XP000590871 ISSN: 0014-2980 the whole document</p>	2,8
A	<p>DANILENKO D M (REPRINT) ET AL: "AGP - 1, a novel member of the tumor necrosis factor family, induces hepatic necrosis and inflammation in transgenic mice." FASEB JOURNAL, (28 FEB 1997) VOL. 11, NO. 3, PP. 2951-2951, XP002141678 the whole document</p>	1-19



**INTERNATIONAL SEARCH REPORT**  
Information on patent family members

International Application No

PCT/US 00/08004

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9701633 A	16-01-1997	AU 5596599 A	13-01-2000
		AU 708239 B	29-07-1999
		AU 6340796 A	30-01-1997
		CA 2225378 A	16-01-1997
		EP 0835305 A	15-04-1998
		JP 11508445 T	27-07-1999
		NO 976045 A	02-03-1998
		NZ 311982 A	30-08-1999
		US 5763223 A	09-06-1998
WO 9746686 A	11-12-1997	AU 3381097 A	05-01-1998
		CA 2256464 A	11-12-1997
		EP 0918860 A	02-06-1999